

- ASHOOR, S.H. & CHU, F.S. (1973b). Inhibition of muscle aldolase by penicillic acid and patulin in vitro. *Food Cosmet. Toxicol.*, **11**: 995-1000.
- AUFFRAY, Y. & BOUTIBONNES, P. (1986). Evaluation of the genotoxic activity of some mycotoxins using *Escherichia coli* in the SOS spot test. *Mutat. Res.*, **171**, 79-82.
- AUFFRAY, Y. & BOUTIBONNES, P. (1987). Genotoxic activity of some mycotoxins using the SOS chromotest- *Mycopathologia*, **100**: 49-53.
- AUFFRAY, Y. & BOUTIBONNES, P. (1988) Induction of SOS function in *Escherichia coli* by some mycotoxins. *Tox. Ass.: An Int. J.*, **3**: 371-378.
- BARTSCH, H., MALAVEILLE, C., CAMUS, A.M., MARTEL-PLANCHE, G., BRUN, G., HAUTEFEUILLE, A., SABADIE, N., BARBIN, A., KUROKI, T., DREVON, C., PICCOLI, C. & MONTESANO, R. (1980). Validation and comparative studies on 180 chemicals with *S. typhimurium* strains and V79 Chinese hamster cells in the presence of various metabolizing systems. *Mutat. Res.*, **76**: 1-50.
- BECCI, P.J., HESS, F.G., JOHNSON, W.D., GALLO, M.A., BABISH, J.G., DAILEY, R.E., & PARENT, R.A. (1981). Long-term carcinogenicity and toxicity studies of patulin in the rat. *J. Appl. Toxicol.*, **1**: 256-261.
- BELITSKY, G.A., Khovanova, E.M., BUDUNOVA, I.V., SHARUPTIS, H.G., (1985) Mycotoxin induction of somatic mosaicism in *Drosophila* and DNA repair in mamalian liver cell cultures. *Cell Biology and Toxicology*, **1**: 133-143
- BOURDIOL, D., ESCOULA, L. & SALVAYRE, R. (1990) Effect of patulin on microbicidal activity of mouse peritoneal macrophages. *Food Chem. Toxic.*, **28**: 29-33.
- BRAUNBERG, R.C., GANTT, O.O. & FRIEDMAN, L. (1982). Toxicological evaluation of compounds found in food using rat renal explants. *Food Chem. Toxicol.*, **20**: 541-546.
- BURGER, M.G., BRAKHAGE, A.A., CREPPY, E.E., DIRHEIMER, G. ROSCHENTHALER, R.J. (1988). Toxicity and mutagenicity of patulin in different test systems. *Arch. Toxicol (suppl)*, **12**: 347-351.
- BURGHARDT, R., BARHOUMI, R., LEWIS, E.H., HARTFORD BAILEY, R., PYLE, K.A., CLEMENT, B.A. & PHILLIPS, T.D. (1992) Patulin-induced cellular toxicity: a vital fluorescense study. *Toxicol. Appl. Pharmacol.*, **112**: 235-244
- CIEGLER, A., BECKWITH, A.C. & JACKSON, L.K. (1976). Teratogenicity of patulin and patulin adducts formed with cysteine. *Appl. Environ. Microbiol.*, **31**: 664-667.
- COORAY, R., KIESSLING, K.H. & LINDAHL-KIESSLING, K. (1982). The effects of patulin and patulin-cysteine mixtures on DNA synthesis and the frequency of sister-chromatid exchanges in human lymphocytes. *Food Chem. Toxicol.*, **20**: 893-898.
- DAILEY, R.E., BLASCHKA, A.M. & BROUWER, E.A. (1977a). Absorption, distribution, and excretion of ¹⁴C-patulin by rats. *J. Toxicol. Environ. Health*, **3**: 479-489.
- DAILEY, R.E., BROUWER, E., BLASCHKA, A.M., REYNALDO, E.F., GREEN, S., MONLUX, W.S. & RUGGLES, D.I. (1977b). Intermediate-duration toxicity study of patulin in rats. *J. Toxicol. Environ. Health*, **2**: 713-725.

- DEVARAJ, H., & DEVARAJ, N., (1987) Rat intestinal lipid changes in patulin toxicity. *Indian J. of Exp. Biol.*, **25**: 637-638.
- DEVARAJ, H., RADHA-SHANMUGASUNDARAM, K. & SHANMUGASUNDARAM, E.R. (1982a). Neurotoxic effect of patulin. *Indian J. Exp. Biol.*, **20**: 230-231. *Health*, **3**: 479-490.
- DEVARAJ, H., RADHA-SHAMMUGASUNDARAM, K. & SHANMUGASUNDARAM, E.R. (1982b). Effect of patulin on intestinal amino acid uptake. *Curr. Sci. (Bangalore)*, **51**: 602-606.
- DEVARAJ, H., SHANMUGASUNDARAM, K.R. & SHANMUGASUNDARAM, E.R.B. (1986). Role of patulin as a diabetogenic lactone. *Indian J. Exp. Biol.*, **24**: 458-459.
- DEVARAJ, H., SUSEELA, R.E., DEVARAJ, N. (1986) Patulin toxicosis in chicks. *Current Science*, **55**: 998-999.
- DICKENS, F. & JONES, H.E.H. (1961). Carcinogenic activity of a series of reactive lactones and related substances. *Br. J. Cancer*, **15**: 85-100.
- DUBECH, N., GUELFY, J.F., ESCOULA, L., (1993) Activité immunomodulatrice de la patuline sur les fonctions des polynucléaires neutrophiles du chien. *Revue Med. Vet.*, **144**: 553-557.
- DULANEY, E.L., & JACOBSEN, C.A. (1987) Synergy of patulin with other antibiotics. *The Journal of Antibiotics*, **40**: 1211-1212
- EPSTEIN, S.S., ARNOLD, E., ANDREA, J., BASS, W. & BISHOP, Y. (1972). Detection of chemical mutagens by the dominant lethal assay in the mouse. *Toxicol. Appl. Pharmacol.*, **23**: 288-325.
- ESCOULA, L., BOURDIOL, D., LINAS, M.D., RECCO, P., & SEGUELA, J.P. (1988a) Enhancing resistance and modulation of humoral immune response to experimental *Candida albicans* infection by patulin. *Mycopathologia*, **103**: 153-156.
- ESCOULA, L., MORE, J. & BARADAT, C. (1977). The toxins of *Byssoschlamys-nivea*. Part I. Acute toxicity of patulin in adult rats and mice. *Ann. Rech. Vet.*, **8**: 41-49.
- ESCOULA, L., THOMSEN, M., BOURDIOL, M., PIPY., B., PEURIERE, S., & ROUBINET, S. (1988b) Patulin immunotoxicology: effect on phagocyte activation and the cellular and humoral immune system of mice and rabbits. *Int. J. Immunopharmac.*, **10**: 983-989
- FUKS-HOLMBERG, D. (1980). The influence of patulin on rat fetus and rat and human placenta. *Toxicol.*, **18**: 437-442.
- GABRIDGE, M.G. & LEGATOR, M.S. (1969). A host-mediated microbiological assay for the detection of mutagenic compounds. *Proc. Soc. Exp. Biol. Med.*, **130**: 831-834.
- GARZA, H.C., SWANSON, B.G. & BRANEN, A.L. (1977). Toxicology study of patulin in monkeys. *J. Food Sci.*, **42**: 1229-1231.
- GOPALAKRISHNAN, V.K. & SAKTHISEKARAN, D (1991) Effect of patulin on albumin fraction of plasma proteins studied in rats. *Biochemistry International*, **25**: 461-475.

GYE, W.E. (1943). Patulin in the common cold. III. Preliminary trial in the common cold. *Lancet* ii: 630-631.

HAYES, A.W. (1977). Effect of patulin on Krebs cycle intermediate-stimulated oxygen consumption. *Tox.Appl.Pharmacol.*, **141**: 165.

HAYES, A.W., PHILLIPS, T.D., WILLIAMS, W.L. & CIEGLER, A. (1979). Acute toxicity of patulin in mice and rats. *Toxicology*, **13**: 91-100.

HINTON, D.M., RILEY, R.T., SHOWKER, J.L. & RIGSBY, W.E. (1989) Patulin-induced ion flux in cultured renal cells and reversal by dithiothreitol and glutathione: a scanning electron microscopy (SEM) X-Ray microanalysis study. *J. Biochem. Toxicology*, **4**: 47-54.

HOPKINS, W.A. (1943). Patulin in the common cold. IV. Biological properties: Extended trial in the common cold. *Lancet* ii: 631-634.

HRADEC, J. & VESELY, D. (1989) The initiator tRNA acceptance assay as a short-term test for carcinogens. 4. Results with 20 mycotoxins. *Carcinogenesis*, **10**: 213-215.

IARC (1976) International Agency for Research on Cancer. Monographs on the evaluation of carcinogenic risk of chemicals to man: *Vol.10* Some naturally occurring substances, 205-210.

IARC (1986) International Agency for Research on Cancer. Monographs on the evaluation of carcinogenic risk of chemicals to man: *Vol.40* Some naturally occurring and synthetic food components,flurocoumarins and ultraviolet radiation, 83-98.

KANGSADALAMPAI, K., SALUNKHE, D.K. & SHARMA, R.P. (1981). Patulin and rubratoxin B: Interactions of toxic and hepatic effects and mutagenic potential. *J. Food Protection*, **44**: 39-42.

KATZMAN, P.A., HAYS, E.E., CAIN, C.K., VAN WYK, J.J., REITHEL, F.J., THAYER, S.A., DOISY, E.A., GABY, W.L., CARROLL, C.J., MUIR, R.D. & JONES, L.R., (1944). Clavacin, an antibiotic substance from *Aspergillus clavatus*. *J. Biol. Chem.*, **154**: 475-486.

KORTE A. (1980). Comparative analysis of chromosomal aberrations and sister chromatid exchanges in bone-marrow cells of Chinese hamsters after treatment with aflatoxin B1, patulin and cyclophosphamide. *Mutat. Res.*, **74**: 164.

KORTE, A. & RUCKERT, G. (1980). Chromosomal analysis in bone-marrow cells of Chinese hamsters after treatment with mycotoxins. *Mutat. Res.*, **78**: 41-49.

KORTE, A., SLACIK-ERBEN, R. & OBE, G. (1979). The influence of ethanol treatment on cytogenetic effects in bone marrow cells of Chinese hamsters by cyclophosphamide, aflatoxin B1 and patulin. *Toxicology*, **12**: 53-61.

KRIVOBOK, S., OLIVIER, P., MARZIN, D.R., SEIGLE-MURANDI, F. & STEIMAN, R. (1987). Study of the genotoxic potential of 17 mycotoxins with the SOS chromotest. *Mutagenesis*, **2**: 433-439.

KUBIAK, R., KOSZ-VNENCHAK, M. (1983). Mutagenic properties of mycotoxins as naturally occurring mutagens: Chromosome aberrations and SCEs induced by patulin. *Mutat.Res.*, **113**: 273.

- KUCZUK, M.H., BENSON, P.M., HEATH, H. & HAYES, A.W. (1978). Evaluation of the mutagenic potential of mycotoxins using *Salmonella typhimurium* and *Saccharomyces cerevisiae*. *Mutat. Res.*, **53**: 11-20.
- LEE, K.S. & ROSCHENTHALER, R.J. (1986). DNA-damaging activity of patulin in *Escherichia coli*. *Appl. Environ. Microbiol.*, **52**: 1046-1054.
- LINDROTH, S. & VON WRIGHT, A. (1978). Comparison of the toxicities of patulin and patulin adducts formed with cysteine. *Appl. Environ. Microbiol.*, **35**: 1003-1007.
- MADHYASTHA, M.S., MARQUARDT, R.R., MASI, A., BORSA, J., & FROHLICH, A.A. (1994) Comparison of toxicity of different mycotoxins to several species of bacteria and yeasts: use of *Bacillus brevis* in a disc diffusion assay. *J. Food Protection*, **57**: 48-53.
- MADIYALAKAN, R. & SHANMUGASUNDARAM, E.R. (1978). Effect of patulin on mouse liver glycogen phosphorylase. *Indian J. Exp. Biol.*, **16**: 1084-1085.
- MAYER, V.W. & LEGATOR, M.S. (1969). Production of petite mutants of *Saccharomyces cerevisiae* by patulin. *J. Agric. Food Chem.*, **17**: 454-456.
- McKINLEY, E.R. & CARLTON, W.W. (1980a). Patulin mycotoxicosis in Swiss ICR mice. *Food Cosmet. Toxicol.*, **18**: 181-187.
- McKINLEY, E.R. & CARLTON, W.W. (1980b). Patulin mycotoxicosis in the Syrian hamster. *Food Cosmet. Toxicol.*, **18**: 173-179.
- McKINLEY, E.R., CARLTON, W. W. & BOON, G.D. (1982). Patulin mycotoxicosis in the rat: Toxicology, pathology and clinical pathology. *Food Chem. Toxicol.*, **20**: 289-300.
- MIURA, S., HASUMI, K., and ENDO, A., (1993) Inhibition of protein prenylation by patulin. *Federation of European Biochemical Societies letters* 12137, **318**: 88-90
- MORI, H., KAWAI, K., OHBAYASHI, F., KUNIYASU, T., YAMAZAKI, M., HAMASAKI, T. & WILLIAMS, G.M. (1984). Genotoxicity of a variety of mycotoxins in the hepatocyte primary culture DNA repair test using rat and mouse hepatocytes *Cancer Res.*, **44**: 2918-2923.
- MOULE, Y. & HATEY, F. (1977). Mechanism of the *in vitro* inhibition of transcription by patulin, a mycotoxin from *Byssoschlamys nivea*. *FEBS Lett.*, **74**: 121-125.
- NISHIE, K., CUTLER, H.G., COLE., R.J. (1989) Toxicity of trichothecenes, moniliformin, Zearalenone/ol, Griseofulvin, Patulin, Pr Toxin and Rubratoxin B on Protozoan *Tetrahymena pyriformis*. *Research Communications in Chemical Pathology and Pharmacology*, **65**: 197-210.
- OSSWALD, H., FRANK, H.K., KOMITOWKSI, D. & WINTER, H. (1978). Long-term testing of patulin administered orally to Sprague-Dawley rats and Swiss mice. *Food Cosmet. Toxicol.*, **16**: 243-247.
- PAUCOD, J.C., KRIVOBOK, S., & VIDAL, D. (1990) Immunotoxicity testing of mycotoxins T-2 and patulin on Balb/C mice. *Acta Microbiol. Hungarica*, **37**: 331-339.
- PHILLIPS T.D. & HAYES, A.W. (1977). Effects of patulin on adenosine triphosphatase in the mouse. *Toxicol. Appl. Pharmacol.*, **42**: 175-187.

- PHILLIPS, T.D. & HAYES, A.W. (1978). Effects of patulin on the kinetics of substrate and cationic ligand activation of adenosine triphosphatase in mouse brain *J. Pharmacol. Exp. Ther.*, **205**: 606-616.
- REDDY, C.S., CHAN, P.K. & HAYES, A.W. (1978). Teratogenic and dominant lethal studies of patulin in mice. *Toxicology*, **11**: 219-223.
- RIHN, B., LUGNIER, A.A.J., DIRHEIMER, G. (1986) Morphological alterations induced by patulin on cultured hepatoma cells. *Arch. Toxicol., Suppl.* **9**: 275-278.
- RILEY, R.T., HINTON, D.M., SHOWKER, J.L., RIGSBY, W., NORRED, W.P. (1990) Chronology of patulin-induced alterations in membrane function of cultured renal cells, LLC-PK₁. *Tox. Appl. Pharm.*, **102**: 128-141
- RILEY, R.T., & SHOWKER, J.L. (1991) The mechanism of patulin's cytotoxicity and the antioxidant activity of indole tetramic acids. *Tox. Appl. Pharm.* **109**: 108-126.
- ROLL, R., MATTHIASCHK, G., KORTE, A. (1990) Embryotoxicity and mutagenicity of mycotoxins. *J. Environ. Pathol. Toxicol. Oncol.*, **10**: 1-7.
- SAKAI, M., ABE, K., OKUMURA, H., KAWAMURA, O., SUGIURA, Y., HORIE, Y., UENO, Y. (1992) Genotoxicity of fungi evaluated by SOS microplate assay. *Natural Toxins*, **1**: 27-34.
- SAKTHISEKARAN, D., & SHANMUGASUNDARAM, E.R. (1990) Effect of patulin on the kinetic properties of the enzyme aldolase studied in rat liver. *Biochemistry International*, **21**: 117-134.
- SAKTHISEKARAN, D., SHANMUGASUNDARAM, K.R., SHANMUGASUNDARAM E.R.B. (1989) Effect of patulin on some enzymes of carbohydrate metabolism studied in rats. *Biochemistry International*, **19**: 37-51.
- SEIGLE-MURANDI, F., STEIMAN, R., KRIVOBOK, S., BERIEL, H. & BENOIT-GUYOD, J.L. (1992) Antitumour activity of patulin and structural analogs. *Pharmazie*, **47**: 288-291
- SHARMA, R.P (1993) Immunotoxicity of mycotoxins. *J. Dairy Sci.*, **76**: 892-897.
- SIRAJ, M.Y. & HAYES, A.W. (1978). induction of hepatic mixed function oxidases by patulin in male mice. *Fed. Proc.*, **37**: 320.
- SMALL, M.H., SMITH, E.E., BRAITHWAITE, C.E., PHILIPS, T.D. & REINE, A.H. (1992) Effects of patulin on postimplantation rat embryos. *Toxicologist*, **12**: 334
- SORENSEN, W.G., GERBERICK, G.F., LEWIS, D.M., Castranova, V., (1986) Toxicity of mycotoxins for the rat pulmonary macrophage *in vitro*. *Env. Health Persp.*, **66**: 45-53.
- SPEIJERS, G.J.A., FRANKEN, M.A. & VAN LEEUWEN F.X. (1988). Subacute toxicity study of patulin in the rat: Effects on the kidney and the gastrointestinal tract. *Food Chem. Toxicol.*, **26**: 23-30.
- SPEIJERS, G.J.A., FRANKEN, M.A.M., VAN LEEUWEN, F.X.R., VAN EGMOND, H.P., BOOT, R., LOEBER, J.G. (1986) Subchronic oral toxicity study of patulin in the rat. Report no. 618314 001. Rijksinstituut voor Volksgezondheid en Milieuhygiëne.

- SPEIJERS, G.J.A., KOLKMAN, R., FRANKEN, M.A.M., VAN LEEUWEN, F.X.R., DANSE, L.H.J.C. (1985) Subacute toxiciteit van patuline in de rat. Rapport nr. 617903 001. Rijksinstituut voor Volksgezondheid en Milieuhygiëne.
- STETINA, R. & VOTAVA, M. (1986). Induction of DNA single-strand breaks and DNA synthesis inhibition by patulin, ochratoxin A, citrinin, and aflatoxin B₁ in cell lines CHO and AWR6. *Folia Biol.*, Prague, **32**: 128-144.
- TASHIRO, F., HIRAL, K. & UENO, Y. (1979). Inhibitory effects of carcinogenic mycotoxins on deoxyribonucleic acid-dependent ribonucleic acid. *Appl. Environ. Microbiol.*, **38**: 191-196.
- THUST, R., KNEIST, S. & MENDEL, J. (1982). Patulin, a further clastogenic mycotoxin, is negative in the SCE assay, in Chinese hamster V79-E cells *in vitro*. *Mutat. Res.*, **103**: 91-97.
- UENO, Y. & KUBOTA, K. (1976). DNA-attacking ability of carcinogenic mycotoxins in recombination-deficient mutant cells of *Bacillus subtilis*. *Cancer Res.*, **38**: 445-451.
- UENO Y., KUBOTA, K., ITO, T. & NAKAMURA, Y. (1978). Mutagenicity of carcinogenic mycotoxins in *Salmonella typhimurium*. *Cancer Res.*, **38**: 536-542.
- UMEDA, M., TSUTSUI, T. & SAITO, M. (1977). Mutagenicity and inducibility of DNA single-strand breaks and chromosome aberrations by various mycotoxins. *Gann*, **68**: 619-625.
- VON WRIGHT, A. & LINDROTH, S. (1978). The lack of mutagenic properties of patulin and patulin adducts formed with cysteine in *Salmonella* test systems. *Mutat. Res.*, **58**: 211-216.
- WEHNER, F.C., THIEL, P.G., VAN RENSBURG, S.J. & DEMASIUS, I.P.C. (1978). Mutagenicity to *Salmonella typhimurium* of some *Aspergillus* and *Penicillium* mycotoxins. *Mutat. Res.*, **58**: 193-203.
- WITHERS, R.F.J. (1966). The action of some lactones and related compounds on human chromosomes. In: Landa, Z. (ed.), *Mechanism of Mutation and Inducing Factors*, Prague Academia, 359-364.
- WURGLER, F.E., FRIEDERICH, U. & SCHLATTER, J. (1991) Lack of mutagenicity of ochratoxin A and B, citrinin, patulin and cneistine in *Salmonella typhimurium* TA102. *Mutation Research*, **261**: 209-216
- YANAGISAWA, K., NISHIO, K., GOTOH, S. (1987) Screening for carcinogens by the DNA synthesis inhibition test using human fibroblasts. *Mutation Research*, **183**: 89-94.

Long-term Carcinogenicity and Toxicity Studies of Patulin in the Rat

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Patulin is a mycotoxin produced by a variety of *Penicillium* and *Aspergillus* species which are likely natural contaminants of various foods. The present study was conducted to determine the effects of lifetime administration of patulin in FDRL Wistar rats. Animals received patulin by gastric intubation three times per week at the level of 0.0, 0.1, 0.5 and 1.5 mg per kg body weight. The animals used in this lifetime study were derived from F₀ parents exposed to equivalent levels of patulin for 4 weeks before mating, and throughout mating, gestation and lactation. Patulin treatment at 0.5 and 1.5 mg kg⁻¹ to male rats caused a significant decrease in body weight gain in comparison to controls. Body weights of treated female rats were similar to that of control rats. No consistent significant differences among groups were noted in the hematology, clinical chemistry or urine analysis parameters measured during or at the termination of the study. Patulin administered to male and female rats at 1.5 mg kg⁻¹ caused a significantly increased mortality rate as compared to respective control animals. The cause of death appeared to be increased pulmonary and laryngotracheal inflammation. No tumorigenic effect of patulin was observed.

INTRODUCTION

Patulin is a water-soluble β -unsaturated lactone, 4-hydroxy-4H-furo(3,2c)pyran-2(6H)-one. The first disease linked to patulin occurred in Japan when dairy cattle died after eating feed artificially contaminated with *Penicillium urticae*.¹ Patulin has been isolated from several species of *Penicillium* and *Aspergillus*,² which are likely natural contaminants of various foodstuffs. The presence of the mycotoxin in different varieties of apples and pears with brown rot was studied; patulin was found in about 50% of the samples investigated³⁻⁵ at levels as high as 1000 ppm. Patulin has also been isolated from flour⁶ and malt feed.¹ Furthermore, the rather widespread occurrence of patulin and its stability in apple juice have been established.⁷⁻⁹

Dickens and Jones¹⁰ reported that patulin, when administered subcutaneously twice weekly to rats for approx. 15 months, produced sarcoma at the injection sites. However, Osswald *et al.*¹¹ found that patulin did not display tumorigenic activity when administered orally to rats for 64 weeks (358 mg per kg body weight, total dose). The acute oral LD₅₀ in rats has been shown to be 32.5 mg kg⁻¹.¹²

The present study was conducted to evaluate the carcinogenicity and toxicity of lifetime administration of patulin in rats for the purpose of determining the safety of patulin as a potential food contaminant.

EXPERIMENTAL

Animals

Wistar rats (FDRL, Wistar derived) 6-8 weeks of age at

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the time of the start of the experiment were individually housed in an environment-controlled room, artificially illuminated for 12 h each day, and maintained at 22 ± 2°C. All animals received Purina Rat Chow (Ralston Purina Company, St Louis, Missouri) and tap water *ad libitum* and were acclimated to their new surroundings for 14 days prior to the start of dosing.

Materials

Patulin was supplied by Makor Chemicals Ltd, Jerusalem, Israel. The patulin used was found to be greater than 95% pure when compared to an ultrapure standard sample and only a single compound was detected when analyzed by UV absorbance, thin layer chromatography and high pressure liquid chromatography. Because the stability of patulin in aqueous solutions is erratic at pH values above 7.0, the crystalline material was dissolved in 1 mM citrate buffer, pH 5.0, immediately prior to each day's dosing.

Design of experiment

The dose levels of patulin used were 0, 0.1, 0.5 and 1.5 mg per kg body weight. These dose levels were selected based on the results of an intermediate duration toxicity study.¹² Test solutions of patulin were administered by gastric intubation 3 times per week (Monday, Wednesday, Friday) except during pregnancy when females were treated 7 days per week. Dosing solutions were adjusted at the end of each week to reflect the changes in body weight. Rats in the control groups were given citrate buffer alone. The temperature of the intubation solution was approx. 22°C.

Groups of 50 rats per sex per level for patulin and 70 rats per sex for the control group were used for the F₀

generation. After 4 weeks of patulin treatment, all rats of the F_0 generation were paired one male to one female within groups to breed an F_1 generation. Female F_0 generation rats received patulin throughout the mating, gestation and lactation phases of the study. After weaning, the F_0 generation male and female rats were killed and necropsies were performed. Tissues were fixed using 10% neutral buffered formalin and grossly abnormal areas were subjected to microscopic examination. Microscopic examinations were conducted on paraffin-embedded, hematoxylin and eosin-stained sections.

From the F_1 generation, 70 rats per sex per level for patulin and 110 rats per sex for the control group were selected for the chronic dosing study. No more than 3 rats per sex from each patulin-treated litter and 4 rats per sex from each control litter were randomly assigned to groups. Animals received their first gastric intubation of patulin when they were approx. 28 days of age; this is considered day 1 of the chronic study. Rats were weighed and food consumption was measured weekly. Animals were observed daily for external signs of patulin toxicity. Blood samples for hematology and clinical chemistry determinations were collected from the periorbital plexus from 10 randomly selected rats per sex per group at 6, 12, 18 and 24 months after the initiation of the chronic study. Total and differential leucocyte count, erythrocyte count, hemoglobin, hematocrit, prothrombin time, urea nitrogen, glutamate oxaloacetate transaminase, glucose, sodium and potassium values were determined. Urine was collected

at the same time periods and measurement of specific gravity and pH as well as microscopic examinations were made.

Five randomly selected rats per sex per group were killed at 6 and 12 months and 10 randomly selected rats per sex per group were killed 18 and 24 months after the initiation of the chronic studies. All surviving male and female animals were killed after 109 weeks of treatment. Complete gross and microscopic necropsy examinations were conducted on all animals used for the chronic study. Tissues were fixed as above. At necropsy, heart, kidneys, liver, ovaries, spleen and testes were weighed. In addition to the organs weighed, the following organs were examined microscopically from paraffin-embedded, hematoxylin and eosin-stained sections: adrenal glands, aorta, brain, cecum, colon, diaphragm, duodenum, epididymides, esophagus, eyes with optic nerve, femur, forestomach, ileum, jejunum, lungs, lymph nodes, mammary gland, pancreas, parathyroid glands, pituitary gland, prostate, rectum, salivary gland, sciatic nerve, seminal vesicles, skeletal muscle, skin, spinal cord, sternum, stomach, thymus, thyroid gland, trachea, urinary bladder, uterus, vagina, and any grossly abnormal tissue.

Statistical analysis

Body weight, food consumption, organ weight and clinical data were evaluated using analysis of variance. Pathology incidence data and indexes of reproduction and lactation were analyzed using a chi-square test with Yate's correction for 2×2 contingency tables.

Table 1. Reproduction and lactation data of rats given patulin

	Patulin level (mg per kg body wt)			
	0.0	0.1	0.5	1.5
Number of females mated	70	50	50	50
Number of dams surviving	70	50	49	40
Number of successful matings	68	47	47	42
Total number of pups produced	868	555	553	459
Number of pups per litter:				
Cast alive	12.5	11.6	11.4	11.9
Cast dead	0.3	0.2	0.6	0.2
Indexes:				
Fertility ^a	97.1	94.0	94.0	84.0
Gestation ^b	100.0	100.0	97.9	90.5
Viability ^c	97.0	97.8	96.2	99.1
Lactation ^d	92.6	90.0	91.5	88.6
Mean body weight (g) per pup:				
at birth	6.2	6.4	6.2	6.4
4 days	10.3	10.5	10.6	10.7
21 days	38.6	39.6	38.2	40.0

^a % matings resulting in pregnancies.

^b % pregnancies resulting in litters cast alive.

^c % pups cast alive that survived to 4 days.

^d % pups alive at 4 days that survived to 21 days.

RESULTS

F_0 generation

A summary of the reproduction and lactation data for the F_0 generation is shown in Table 1. No statistically significant differences were noted among groups in the parameters measured. Body weight gain by F_0 generation male and female rats showed no significant differences among groups. Histopathologic evaluation of grossly abnormal tissues from F_0 generation animals revealed no effect which could be attributed to patulin treatment.

F_1 generation

Body weight data. The group mean body weights of the F_1 generation animals are shown in Table 2. A significant reduction in body weight gain was noted for male rats at the 0.5 and 1.5 mg kg⁻¹ dose levels ($p \leq 0.05$). No significant differences in body weight gain of female rats was noted among groups.

Food consumption data. Intermittent significant differences in food consumption of male and female rats were noted among groups; however, no consistent or significant pattern was observed.

Table 2. Group mean body weight of rats given patulin

Sex and patulin level (mg per kg body wt per day)	Body weight (g) \pm SE at week										
	1 ^a	10	20	30	40	50	60	70	80	90	100
Male:											
0	62 \pm 1	354 \pm 3	425 \pm 4	463 \pm 6	499 \pm 6	524 \pm 6	548 \pm 7	549 \pm 9	548 \pm 10	541 \pm 14	543 \pm 13
0.1	60 \pm 1	352 \pm 7	419 \pm 5	465 \pm 6	493 \pm 6	516 \pm 7	541 \pm 10	549 \pm 11	523 \pm 17	549 \pm 19	530 \pm 22
0.5	60 \pm 1	349 \pm 4	413 \pm 5	446 \pm 6	470 \pm 7 ^c	492 \pm 8 ^c	512 \pm 9 ^c	520 \pm 12	514 \pm 16 ^c	514 \pm 19 ^c	499 \pm 23 ^c
1.5	63 \pm 1	349 \pm 4	412 \pm 6	436 \pm 7 ^c	455 \pm 7 ^c	471 \pm 9 ^c	487 \pm 8 ^c	485 \pm 13 ^c	458 \pm 11 ^c	b	b
Female:											
0	58 \pm 1	217 \pm 2	252 \pm 2	264 \pm 3	280 \pm 3	296 \pm 4	315 \pm 4	330 \pm 5	340 \pm 7	346 \pm 7	348 \pm 7
0.1	58 \pm 1	220 \pm 2	254 \pm 3	265 \pm 4	279 \pm 4	297 \pm 5	314 \pm 6	331 \pm 8	343 \pm 10	347 \pm 10	355 \pm 15
0.5	56 \pm 1	214 \pm 2	245 \pm 3	256 \pm 3	269 \pm 4	286 \pm 4	304 \pm 6	322 \pm 8	324 \pm 10	330 \pm 10	344 \pm 12
1.5	58 \pm 1	221 \pm 2	254 \pm 2	266 \pm 3	274 \pm 4	292 \pm 5	306 \pm 6	322 \pm 8	317 \pm 10	326 \pm 11	340 \pm 14

^a Animals were approx. 28 days of age at the start of the chronic study (week 1).

^b High dose male group terminated at 83 weeks.

^c Significantly different from respective control, $p < 0.05$.

Hematology and urine analysis. Hematology data from F₁ generation male animals sampled at 12, 18 and 24 months showed no significant differences among groups, nor were there differences among groups for female animals sampled at 6, 12 and 18 months. At 6 months, a significant decrease in the erythrocyte count was noted in the low and high dose male animals when compared to controls (Table 3). At 24 months, a significant decrease in the leucocyte count was noted on female animals at the middle dose level in comparison to control rats (Table 3). It is important to note that all the mean values in the above groups were within the normal physiological range for rats.

No significant differences were noted in any urine parameters measured throughout the course of the study.

Clinical chemistry. Data from F₁ generation male and female animals sampled at 12, 18 and 24 months showed no toxicologically significant differences among groups. An

apparent dose-related decrease in sodium and potassium levels of male animals given patulin was noted at 6 months (Table 4). In female rats sampled at 6 months, an apparent dose-related increase in urea nitrogen levels was noted (Table 4). All sodium, potassium and urea nitrogen values obtained through the course of the study were within the normal range for rats.

Organ weights. Data taken at 6, 12, 18 and 24 months are shown in Tables 5 and 6.

At 6 months, absolute but not relative heart weights of the high dose female rats were significantly increased in comparison to control rats ($p \leq 0.05$). Relative and absolute ovary weights were significantly increased in the high dose group in comparison to control rats ($p \leq 0.05$) at 12 months but were comparable to controls at other time periods. At 18 months, the absolute but not relative liver and spleen weights of the middle and high dose male

Table 3. Summary of hematology data of rats given patulin^a

Sex and patulin level (mg per kg body wt)	Erythrocyte count ($\times 10^6 \text{ mm}^{-3}$)	Leucocyte count ($\times 10^3 \text{ mm}^{-3}$)	Hemoglobin (g 100 ml ⁻¹)	Hematocrit (%)
6 months				
Male:				
0	7.8 \pm 0.1	9.4 \pm 0.4	15.4 \pm 0.3	38 \pm 1
0.1	6.9 \pm 0.2 ^b	8.0 \pm 0.5	14.5 \pm 0.5	35 \pm 1
0.5	7.4 \pm 0.2	8.4 \pm 0.4	15.2 \pm 0.4	37 \pm 1
1.5	6.8 \pm 0.2 ^b	7.9 \pm 0.4	15.1 \pm 0.6	34 \pm 2
24 months				
Female:				
0	8.1 \pm 0.6	7.0 \pm 0.3	14.9 \pm 1.0	42 \pm 3
0.1	7.7 \pm 0.2	7.2 \pm 0.5	14.7 \pm 0.4	41 \pm 1
0.5	7.9 \pm 0.2	5.4 \pm 0.4 ^b	15.0 \pm 0.3	42 \pm 1
1.5	7.5 \pm 0.5	5.8 \pm 0.4	14.5 \pm 0.6	40 \pm 2

^a Values are means \pm SE of 10 rats per group.

^b Significantly different from respective control, $p < 0.05$.

Table 4. Summary of clinical chemistry data of rats given patulin^a

Sex and patulin level (mg per kg body wt)	Urea nitrogen (mg 100 ml ⁻¹)	Glucose (mg 100 ml ⁻¹)	SGOT (RF units)	Sodium (meg l ⁻¹)	Potassium (meg l ⁻¹)
6 months					
Male:					
0	20±1	84±5	58±4	163±2	6.9±0.2
0.1	18±1	92±3	63±5	147±3 ^b	5.6±0.2 ^b
0.5	20±1	80±8	57±4	152±3 ^b	5.3±0.2 ^b
1.5	20±1	91±3	60±4	146±2 ^b	4.9±0.2 ^b
Female:					
0	20±1	85±5	63±4	155±4	6.0±0.3
0.1	22±1	94±5	67±6	145±2	4.9±0.2
0.5	23±1 ^b	89±2	56±2	145±3	5.4±0.2
1.5	24±1 ^b	92±3	65±5	146±4	5.5±0.3

^a Values are means ± SE of 10 rats per group.

^b Significantly different from respective control, $p < 0.05$.

Table 5. Absolute organ weights of male rats given patulin

Patulin level (mg per kg body wt)	No. of rats	Group mean ± SE organ weights (g)					Terminal body weight
		Liver	Spleen	Kidneys	Testes	Heart	
6 months							
0	5	17.6±0.3	0.78±0.07	3.1±0.2	3.5±0.1	1.4±0.1	448±13
0.1	5	14.2±0.4	0.73±0.08	2.4±0.4	3.2±0.1	1.2±0.1	427±8
0.5	5	16.5±1.2	0.66±0.04	3.0±0.1	3.5±0.1	1.2±0.1	466±25
1.5	5	16.5±0.8	0.71±0.07	2.2±0.6	3.6±0.1	1.3±0.1	458±10
12 months							
0	5	16.2±0.4	0.80±0.03	3.1±0.1	3.3±0.1	1.6±0.1	524±18
0.1	5	15.6±1.2	0.75±0.04	3.1±0.2	3.3±0.1	1.5±0.1	470±9
0.5	5	15.6±2.2	0.73±0.06	3.1±0.3	3.4±0.2	1.6±0.1	439±29
1.5	5	14.7±1.7	0.72±0.07	3.2±0.1	2.9±0.4	1.7±0.1	459±34
18 months							
0	10	19.6±1.9	1.07±0.08	3.8±0.3	3.3±0.2	1.8±0.1	539±29
0.1	10	17.9±0.6	0.92±0.05	3.7±0.1	3.1±0.2	1.8±0.1	547±26
0.5	10	14.9±0.6 ^a	0.77±0.03 ^a	3.3±0.1	3.3±0.1	1.7±0.1	481±25
1.5	10	14.3±1.2 ^a	0.77±0.09 ^a	3.3±0.2	3.1±0.2	1.7±0.1	444±32
24 months ^b							
0	10	16.0±1.1	0.84±0.08	3.7±0.2	2.8±0.3	1.9±0.1	408±33
0.1	10	15.2±1.1	1.20±0.11	4.2±0.5	3.1±0.2	1.8±0.1	540±34
0.5	10	15.9±0.9	1.09±0.07	4.1±0.4	2.9±0.2	1.8±0.1	463±25

^a Significantly different from respective control, $p < 0.05$.

^b The high dose group was terminated at 83 weeks.

rats were significantly less than that of control animals ($p \leq 0.05$). Spleen weights were significantly increased in the middle dose male rats and the high dose female rats in comparison to controls ($p \leq 0.05$) at 24 months.

Mortality and histopathology. Mortality curves of rats are shown in Fig. 1. Data were analyzed using 2×2 contingency table with Yate's correction and evaluated either the number of animals that survived through a particular time period or the number of animals that died and were sacrificed moribund through the same time period. Similar

results were obtained when the data were analyzed using either parameter. The high dose level of patulin caused a significant increase in the mortality rate in both sexes over the 24-month test period ($p \leq 0.05$). The effect was most pronounced during the first 12 months of the study. The cause of death appeared to be increased pulmonary and laryngotracheal inflammation with production of mucofibrinous exudate that apparently obstructed the tracheal lumen.

The same spectrum of tumors observed historically in rats of this strain was noted in the rats of the present

Table 6. Absolute organ weights of female rats given patulin

Patulin level (mg per kg body wt)	No. of rats	Group mean \pm SE organ weights (g)					Terminal body weight
		Liver	Spleen	Kidneys	Ovaries	Heart	
6 months							
0	5	8.4 \pm 0.6	0.46 \pm 0.03	1.7 \pm 0.1	0.09 \pm 0.01	0.9 \pm 0.02	235 \pm 6
0.1	5	8.3 \pm 0.4	0.55 \pm 0.05	1.7 \pm 0.1	0.09 \pm 0.01	0.9 \pm 0.05	265 \pm 14 ^a
0.5	5	8.0 \pm 0.4	0.46 \pm 0.02	1.8 \pm 0.1	0.10 \pm 0.01	0.9 \pm 0.01	242 \pm 4
1.5	5	9.2 \pm 0.5	0.53 \pm 0.02	1.9 \pm 0.1	0.10 \pm 0.01	1.0 \pm 0.04 ^a	265 \pm 2 ^a
12 months							
0	5	8.1 \pm 0.7	0.49 \pm 0.02	2.0 \pm 0.1	0.09 \pm 0.01	1.1 \pm 0.07	295 \pm 35
0.1	5	8.5 \pm 0.5	0.62 \pm 0.07	2.4 \pm 0.2	0.12 \pm 0.02	1.2 \pm 0.14	289 \pm 15
0.5	5	7.8 \pm 0.7	0.50 \pm 0.03	2.2 \pm 0.1	0.08 \pm 0.01	1.4 \pm 0.07	274 \pm 20
1.5	5	9.4 \pm 0.5	0.60 \pm 0.05	2.2 \pm 0.1	0.16 \pm 0.02 ^a	1.2 \pm 0.02	296 \pm 19
18 months							
0	10	9.5 \pm 0.7	0.63 \pm 0.07	2.5 \pm 0.1	0.13 \pm 0.02	1.3 \pm 0.05	309 \pm 19
0.1	10	9.7 \pm 0.3	0.65 \pm 0.03	2.4 \pm 0.1	0.17 \pm 0.06	1.2 \pm 0.05	325 \pm 23
0.5	10	9.1 \pm 0.4	0.60 \pm 0.04	2.3 \pm 0.1	0.12 \pm 0.01	1.3 \pm 0.06	336 \pm 24
1.5	10	9.6 \pm 0.4	0.55 \pm 0.03	2.2 \pm 0.1	0.12 \pm 0.01	1.3 \pm 0.05	340 \pm 22
24 months							
0	10	11.4 \pm 1.1	0.71 \pm 0.09	2.5 \pm 0.1	0.14 \pm 0.03	1.5 \pm 0.10	325 \pm 20
0.1	10	11.1 \pm 0.8	0.69 \pm 0.02	2.5 \pm 0.1	0.11 \pm 0.01	1.5 \pm 0.08	356 \pm 10
0.5	10	10.0 \pm 0.5	0.66 \pm 0.04	2.4 \pm 0.1	0.15 \pm 0.05	1.4 \pm 0.06	299 \pm 17
1.5	10	10.1 \pm 0.5	1.04 \pm 0.13 ^a	2.5 \pm 0.1	0.11 \pm 0.01	1.5 \pm 0.07	350 \pm 18

^a Significantly different from respective control, $p < 0.05$.

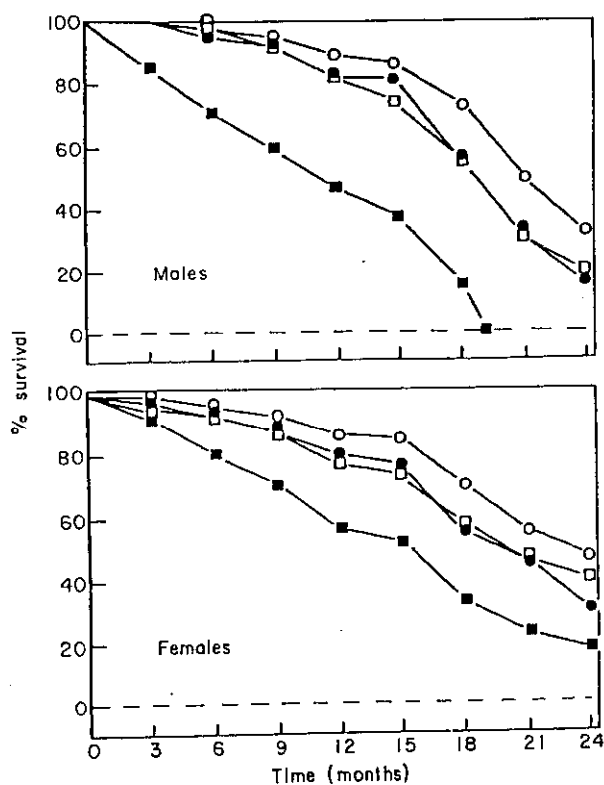


Figure 1. Survival of rats administered patulin by gavage at a level of 0.0 (○), 0.1 (□), 0.5 (●) and 1.5 (■) mg per kg body weight 3 times per week.

study. Statistical analysis of these data showed that the incidence of each type of tumor in any of the patulin-treated groups was comparable to that in the control group. The total numbers of tumors per group, the average numbers per rat and the times of observation of tumors were not affected by any of the levels of treatment. These data in each case were comparable to those of the control group.

DISCUSSION

Patulin, when administered subcutaneously to rats for approx. 15 months, produced sarcomas at the injection sites.¹⁰ However, the validity of the subcutaneous route of administration and the subsequent conclusions drawn relative to carcinogenicity are questionable. Furthermore, patulin did not display tumorigenic activity when administered orally to rats for 64 weeks.¹¹ Because of the relatively short duration of the oral dosing study¹¹ and the questionable significance of the subcutaneous study,¹⁰ the present study was conducted to evaluate the carcinogenicity of patulin when administered to rats for approx. 109 weeks. In our study, no tumorigenic effect of patulin was noted. Analysis of the data showed that the incidence of each type of tumor in any of the patulin-treated groups was comparable to the control group.

Patulin at the high dose level caused a significant increase in the mortality rate in both sexes at the 109-week

test period. The effect was most pronounced during the first year of the study. The major cause of mortality in the patulin-treated group appeared to be pulmonary and laryngotracheal inflammation.

It is not clear, however, whether this effect resulted from systemic toxicity or from the mechanics of multiple intubations. Osswald *et al.*¹¹ have attributed similar findings to dosing methodology. In short-term studies,¹⁷ atelectasis, alveolar septal congestion and intraalveolar hemorrhage were noted in both rats and mice treated with patulin by the intraperitoneal route of administration.

In both rats and mice, patulin has been reported to induce edema of the lungs and brain, visceral organ congestion, and hepatic and renal necrosis.^{13,14} In a two-generation toxicity study of patulin in rats, the only lesion observed was gaseous distention of the gastrointestinal tract at levels greater than 1.5 mg kg⁻¹.¹² In chickens, the principal gross lesion related to patulin was extensive intestinal hemorrhage.¹⁵ In the above studies, the gastrointestinal lesions can probably be attributed to patulin's antibiotic activity resulting in changes in the bacterial flora of the intestinal tract; patulin, as an antibiotic, inhibits Gram-positive

organisms.¹⁶ In the present study, none of these lesions were noted as test effects.

Although various transient differences were noted in hematology and clinical chemistry values of patulin-treated rats when compared to control rats, none was considered treatment-related or of toxicological significance. Similarly, comparisons of organ weight data did not demonstrate treatment-related effects.

Microscopic examination of tissues and organs other than the respiratory tract did not reveal any treatment-related effects. Lesions observed in patulin-treated animals were comparable to those found in control animals and in historical control animals.

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REFERENCES

1. T. Ukai, U. Yamamoto and T. J. Yamamoto, Studies on the poisonous substance from a strain of *Penicillium* (Hori-Yamamoto strain). II. Culture method of Hori-Yamamoto strain and chemical structure of its poisonous substance. *Proc. Pharm. Soc. Jpn* 74, 450 (1954).
2. E. P. Abraham and H. W. Florey, Substances produced by Fungi Imperfecti and Ascomycetes. In *Antibiotics*, ed. by H. W. Florey, E. Chain, N. G. Heatley, M. A. Jennings, A. G. Sanders, E. P. Abraham and M. E. Florey, Vol. 1, p. 273. Oxford University Press, London (1949).
3. H. K. Frank, R. U. Orth and R. Hermann, Patulin in Lebensmitteln pflanzlicher Herkunft. I. Kernobst und daraus hergestellte Produkte. *Z. Lebensm.-Unters. Forsch.* 162, 149 (1976).
4. J. Harwig, Y. K. Chen, B. P. C. Kennedy and P. M. Scott, Occurrence of patulin and patulin-producing strains of *Penicillium expansum* in natural rot of apples in Canada. *Can. Inst. Food Technol. J.* 6, 22 (1973).
5. P. W. Brain, G. W. Wilson and D. Lowe, Production of patulin in apple fruits by *Penicillium expansum*. *Nature (London)* 178, 263 (1956).
6. C. W. Hesseltine and R. R. Graves, Microbiology of flours. *Econ. Bot.* 20, 156 (1966).
7. A. E. Pohland and R. Allen, Stability studies with patulin. *J. Assoc. Off. Anal. Chem.* 53, 688 (1970).
8. P. M. Scott, W. F. Miles, P. Toft and J. G. Dube, Occurrence of patulin in apple juice. *J. Agric. Food Chem.* 20, 450 (1972).
9. P. M. Scott and E. Somers, Stability of patulin and penicillic acid in fruit juices and flour. *J. Agric. Food Chem.* 16, 483 (1968).
10. F. Dickens and H. E. H. Jones, Carcinogenic activity of a series of reactive lactones and related substances. *Br. J. Cancer* 15, 85 (1961).
11. H. Osswald, H. K. Frank, D. Komitowski and H. Winter, Long-term testing of patulin administered orally to Sprague-Dawley rats and Swiss mice. *Food Cosmet. Toxicol.* 16, 243 (1978).
12. R. E. Dailey, E. Brouwer, A. M. Blaschka, E. F. Reynaldo, S. Green, W. S. Monlux and D. I. Ruggles, Intermediate-duration toxicity study of patulin in rats. *J. Toxicol. Environ. Health* 2, 713 (1977).
13. A. W. Hayes, T. D. Phillips, W. L. Williams and A. Ciegler, Acute toxicity of patulin in mice and rats. *Toxicology* 13, 91 (1979).
14. W. A. Broom, E. Bulbring, C. J. Chapman, J. W. F. Hampton, A. M. Thomson, J. Ungar, R. Wein and G. Woolfe, The pharmacology of patulin. *Br. J. Exp. Pathol.* 25, 195 (1944).
15. L. Esconla, J. More and C. Baradat, The toxins of *Byssoschlamys nivea* Westling. 1. Acute toxicity in adult rats and mice. *Ann. Rech. Vet.* 8, 41 (1972).
16. J. Lovett, Patulin toxicosis in poultry. *Poult. Sci.* 51, 2097 (1972).
17. J. Singh, Patulin. In *Antibiotics*, ed. by D. Gottlieb and P. D. Shaw. Vol. 1, p. 621. Springer-Verlag, New York (1967).

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