

45. Esmonde TFG, Will RG, Slattey JM, Knight R, Harries-Jones R, DeSilva R, Matthews WB. Creutzfeldt-Jakob disease and blood transfusion. *Lancet* 1993;**341**:205-7. Links

46. Van Duyn CM, Delasnerie-Laupretre N, Masullo C, Zerr I, Silva R de, Wientjes DPWM, Brandel J-P, Weber T, Bonavita V, Zaidier M, Alperovitch A, Poser S, Granieri E, Hofman A, Will RG. Case-control study of risk factors of Creutzfeldt-Jakob disease in Europe during 1993-1995. *Lancet* 1998;**353**:1081-5. Links

47. Collins S, Law MG, Fletcher A, Boyd A, Kaldor J, Masters CL. Surgical treatment and risk of sporadic Creutzfeldt-Jakob disease: a case-control study. *Lancet* 1999;**353**:693-7. Links

48. Llewelyn CA, Hewitt PA, Knight RSG, Amar K, Cousens S, Mackenzie J, Will RG. Possible transmission of variant Creutzfeldt-Jakob disease by blood transfusion. *Lancet* 2004;**363**:417-21. Links

49. Peren AH, Head MW, Ritchie DL, Bell JE, Ironside JW. Preclinical vCJD after blood transfusion in a PRNP codon 129 heterozygous patient. *Lancet* 2004;**364**:527-9. Links

50. Health Protection Agency. New case of transfusion-associated variant-CJD. *CDR Wkly* 2006;**16**:2-3. Links

51. White SJ, Pal S, Siddique D, Hyare H, Macfarlane R, Joiner S, Linehan JM, Brandner S, Wadsworth JD, Hewitt P, Collinge J. Clinical presentation and pre-mortem diagnosis of blood transfusion-associated variant CJD. *Lancet* 2008;**368**:2061-7. Links

52. Editorial team. Fourth case of transfusion-associated vCJD infection in the United Kingdom. *Euro Surveill* 2007;**12**:E070118.4. Links

53. Brown P, Proeze M, Brandel JP, Sato T, McShane L, Zerr I, Fletcher A, Will RG, Pocchiarri M, Cashman NR, d'Aignaux JH, Cervenakova L, Fradkin J, Schonberger LB, Collins SJ. Iatrogenic Creutzfeldt-Jakob disease at the millennium. *Neurology* 2000;**55**:1075-81. Links

54. Johnston R. Prion diseases. *Lancet Neurol* 2005;**4**:635-42. Links

55. Wilson K, Ricketts MN. A third episode of transfusion-derived vCJD. *Lancet* 2006;**368**:2037-9. Links

56. Prowse CV, Bailey A. Validation of prion removal by leucocyte-depleting filters: a cautionary tale. *Vox Sang* 2000;**79**:248. Links

57. Gregori L, McCombie N, Palmer D, Birch P, Sowemimo-Coker SO, Gulivi A, Rohwer RG. Effectiveness of leucoreduction for removal of infectivity of transmissible spongiform encephalopathies from blood. *Lancet* 2004;**364**:529-31. Links

58. Lucham CA, Turner ML. Managing the risk of transmission of variant Creutzfeldt-Jakob disease by blood products. *Br J Haematol* 2005;**132**:13-24. Links

59. Prowse C. Controlling the blood-borne spread of human prion disease. *ISBT Sci Ser* 2006;**1**:21-4. Links

60. Circulaire N° DGS-SDSC-DHOS/2605/433 du 23 septembre 2005 relative aux recommandations pour le traitement des dispositifs médicaux utilisés chez les sujets ayant reçu des produits sanguins labiles (PSL) provenant de donneurs rétrospectivement atteints de variant de la maladie de Creutzfeldt-Jakob (vMCJ). *Bull Officiel Santé* 2005;no° 16.

61. Wilson K, Ricketts MN. Transfusion transmission of vCJD: a crisis averted. *Lancet* 2004;**364**:477-9. Links

62. Murphy EL, Connor D, McEvoy P, Hirschler N, Busch MP, Roberts P, Nguyen KA, Reich P. Estimating blood donor loss due to the variant CJD travel deferral. *Transfusion* 2004;**44**:645-50. Links

63. Farrugia A, Ironside JW, Gangrande P. Variant Creutzfeldt-Jakob disease transmission by plasma products: assessing and communicating risk in an era of scientific uncertainty. *Vox Sang* 2005;**89**:186-92. Links

64. Wilson K, Ricketts MN. The success of precaution? Managing the risk of transfusion transmission of variant Creutzfeldt-Jakob disease. *Transfusion* 2004;**44**:1475-8. Links

65. Wilson K, Wilson M, Hebert PC, Graham I. The application of the precautionary principle to the blood system: the Canadian blood system's vCJD donor deferral policy. *Transfus Med Rev* 2003;**17**:89-94. Links

66. O'Brien S, Chiavetta JA, Goleman M, Fan W, Nair RC, Sher GD, Vamvakas EC. Predictive ability of sequential surveys in determining donor loss from increasingly stringent variant Creutzfeldt-Jakob disease deferral policies. *Transfusion* 2006;**46**:461-8. Links

67. Martin M, Legras JH, Pouchot E, Trouvin JH. Evaluation du risque transfusionnel vis-à-vis de la variante de la maladie de Creutzfeldt-Jakob en France. *Transfus Clin Biol* 2006;**13**:298-303. Links

68. Houston F, McCutcheon S, Goldmann W, Chong A, Foster J, Siso S, Gonzalez L, Jeffrey M, Hunter N. Prion diseases are efficiently transmitted by blood transfusion in sheep. *Blood* 2008; July 22. [Epub ahead of print]. Links

69. Docra SA, Bennett PG. vCJD and blood transfusion: risk assessment in the United Kingdom. *Transfus Clin Biol* 2006;**13**:307-11. Links

70. Sise S, Gonzalez L, Houston F, Hunter N, Martin S, Jeffrey M. The neuropathologic phenotype of experimental ovine BSE is maintained after blood transfusion. *Blood* 2006;**108**:745-8. Links

71. Brown P, Cervenakova L, McShane LM, Barber P, Rubenstein R, Drohan WN. Further studies of blood infectivity

in an experimental model of transmissible spongiform encephalopathy, with an emphasis on the infectivity of plasma components do not transmit Creutzfeldt-Jakob disease in humans. *Transfusion* 2009;**49**:101-10. Links

72. Brown P. Blood infectivity, processing and screening tests in transmissible spongiform encephalopathy. *Vox Sang* 2005;**89**:63-70. Links

73. Holada K, Vostal JG, Theisen PW, MacSulay C, Gregori L, Rohwer RS. Scrapie infectivity is associated with platelets. *J Virol* 2002;**76**:4649-50. Links

74. Prowse C. Prion removal with filters. *ISBT Sci Ser* 2006;**1**:193-7. Links

75. Sowemimo-Coker S, Kascsak R, Kim A, Andrade F, Pesci S, Kascsak RB, Moskova J, Drohan WN. Exogenous (spiked) and endogenous prion infectivity from red cells with a novel platelet-derived prion filter. *Transfusion* 2005;**45**:1839-44. Links

76. Sowemimo-Coker SO, Pesci S, Andrade F, Kim A, Kascsak RB, Moskova J, Drohan WN. Prion affinity prion-reduction filter removes exogenous infectious prions and endogenous prions from red cell concentrates. *Vox Sang* 2006;**90**:265-75. Links

77. Cervia JS, Sowemimo-Coker SO, Ortolano GA, Wilkins JG, Soriano J, Wadsworth JD, Collinge J. Prion and the role of blood filtration in reducing the risk of transmissible spongiform encephalopathy. *Transfus Med Rev* 2006;**20**:190-206. Links

78. Gregori L, Lambert BC, Gurgel PV, Georgiadis E, Edwardson P, Lathrop JT, MacGregor IR, Hammond D, Rohwer RG. Reduction of transmissible spongiform encephalopathy infectivity of blood cells with prion protein affinity ligands. *Transfusion* 2007;**47**:112-61. Links

79. Gregori L, Gurgel P, Lathrop JT, Edwardson P, Lambert BC, Cardinali PG, Burton DR, Rohwer RG. Reduction in infectivity of endogenous transmissible spongiform encephalopathy prions by adsorption to selective affinity resins. *Lancet* 2006;**368**:2220-00. Links

80. Turner ML. Prion reduction filters. *Lancet* 2006;**368**:2190-1. Links

81. Silveira JR, Raymond GJ, Hughson AG, Race RE, Sim VL, Hayes SP, Caughey B. Prion protein aggregates. *Nature* 2005;**437**:257-61. Links

82. Dodd R. Bovine spongiform encephalopathy, variant CJD, and blood transfusion: a review. *Transfus Med Rev* 2004;**44**:628-30. Links

83. Zerr I, Bodemer M, Gafelier O, Otto M, Poser S, Willfang J, Willrich D, Krutzschmann A, Willrich D, 14-3-3 protein in the cerebrospinal fluid supports the diagnosis of Creutzfeldt-Jakob disease. *Lancet* 1998;**43**:32-40. Links

84. Otto M, Willfang J, Schulz E, Zerr I, Otto A, Prähner A, Glatzer C, Jurek W, Glatzer C, Willrich D, Poser S. Diagnosis of Creutzfeldt-Jakob disease by measurement of 14-3-3 protein in cerebrospinal fluid: a control study. *BMJ* 1998;**316**:577-82. Links

85. Miele G, Manson J, Clinton M. A novel erythrocyte-specific marker for variant Creutzfeldt-Jakob disease. *Nature* 2001;**7**:361-4. Links

86. Barnard G, Helmick B, Maden S, Gibourne C, Patel A. The modification of amyloid fibrils by differential extraction and Delfia as a diagnostic test for BSE in sheep and cattle. *Transfus Med Rev* 2002;**16**:10-14. Links

87. Safar JG, Scott M, Monahan J, Deering C, Didorenko S, Sattavat M, Sanchez M, Serban H, Groth D, Burton DR, Prusiner SB, Wadsworth JW. Diagnosis of variant Creutzfeldt-Jakob encephalopathy or chronic wasting disease by immunoblotting for proteinase-resistant PrP<sup>Sc</sup>. *Transfusion* 2002;**20**:1147-50. Links

88. Ballon A, Seyfort-Brandt W, Lang H, Baron H, Vey W. Immunoblotting for PrP<sup>Sc</sup> in sheep: suitability for human prion detection with enhanced sensitivity. *J Clin Microbiol* 2005;**43**:100-104. Links

89. Schmerr MJ, Jenny AL, Bolgin MS, Miller JM, Farnham AN, Chhabra R, Fooka A, Drohan WN and fluorescent labelled peptide to detect the abnormal prion protein PrP<sup>Sc</sup> associated with a transmissible spongiform encephalopathy. *J Chromatogr B* 2004;**813**:201-206. Links

90. Safar JG, Geschwind MD, Deering C, Didorenko S, Sattavat M, Sanchez M, Serban H, Groth D, Miller BL, Dearmond SJ, Prusiner SB. Diagnosis of human prion disease. *Proc Natl Acad Sci USA* 2005;**102**:3501-6. Links

91. Brown P, Cervenakova L. The modern landscape of transfusion-related prion disease and blood screening tests. *Curr Opin Hematol* 2004;**11**:351-6. Links

92. MacGregor IR. Screening assays for transmissible spongiform encephalopathies in transfused blood. *Transfus Med Rev* 2006;**20**:177-84. Links

93. Minor PD. Technical aspects of the development and validation of tests for variant Creutzfeldt-Jakob disease. *Vox Sang* 2004;**86**:164-75. Links

94. Cooper JK, Ladhani K, Minor D. Reference materials for the development of variant Creutzfeldt-Jakob disease diagnostic assays. *Vox Sang* 2007;**92**:322-10. Links

95. Fajge T, Barclay GR, MacGregor I, Head M, Ironside J, Turner M. Variation in concentration of prion protein in the peripheral blood of patients with variant and sporadic Creutzfeldt-Jakob disease detected by dissociation enhanced lanthanide fluorimmunoassay and flow cytometry. *Transfusion* 2005;**45**:504-13. Links
96. Turner ML. Transfusion safety with regards to prions: ethical, legal and societal considerations. *Transfus Clin Biol* 2007;**13**:317-9. Links
97. Lefrère JJ. The BOTIA project ("Blood and Organ Transmissible Infectious Agents"): a European collection of blood samples and an observatory of agents transmitted by blood transfusion or organ transplantation. *Transfus Clin Biol* 2005;**12**:93-4. Links
98. Vaieron AJ, Boelle PY, Chatignoux E, Cesbron JY. Can a second wave of new variant of the CJD be discarded in absence of observation of clinical non Met-Met cases? *Rev Epidemiol Sante Publique* 2006;**54**:111-5. Links
99. Dietz K, Raddatz G, Wallis J, Müller N, Zerr I, Duerr HP, Lefèvre H, Seifried E, Löwer J. Blood transfusion and spread of variant Creutzfeldt-Jakob disease. *Emerg Infect Dis* 2007;**13**:89-96. Links
100. Hartemann P et le Comité européen SCENHIR. Actualités sur le risque iatrogène d'infection par agent à transmission non conventionnelle lors de la transfusion sanguine et d'un acte invasif. *Hygiène* 2006;**14**:417-22. Links
101. Egin RP, Murphy WG. Beyond leukodepletion: removing infectious prions by filtration. *Transfusion* 2005;**45**:1036-8. Links
102. Wabbot N, Turner M. Prions and the blood and immune systems. *Haematologica* 2005;**90**:542-8. Links

## An update on the assessment and management of the risk of transmission of variant Creutzfeldt-Jakob disease by blood and plasma products

Marc L. Turner and Christopher A. Ludlam

Scottish National Blood Transfusion Service and Department of Haematology, Royal Infirmary, Edinburgh, UK

### Summary

There have been four highly probable instances of variant Creutzfeldt-Jakob disease (vCJD) transmission by non-leucocyte depleted red cell concentrates and it is now clear that the infectious agent is transmissible by blood components. To date there is no reported evidence that the infectious agent has been transmitted by fractionated plasma products, e.g. factor VIII concentrate. This review outlines current and potential risk management strategies including donor deferral criteria, the potential for donor screening, blood component processing and prion reduction filters, plasma product manufacture and the difficulties in identification and notification of those considered 'at risk of vCJD for public health purposes'.

**Keywords:** Creutzfeldt-Jakob disease, blood, plasma products.

This review offers an update on our recent assessment and management of the risk of transmission of variant Creutzfeldt-Jakob disease (vCJD) by blood components and plasma products (Ludlam & Turner, 2005). As that review surveyed perceptions on the nature of the prion agent, the spectrum of prion diseases in animals and man, and the range of animal studies relating to pathogenicity and infectivity (much of which still represents the current level of knowledge), these topics are not reviewed again here, other than where significant new relevant studies have been published. This current review focuses on the state of the art in relation to the safety of blood components and plasma products, which has also been reviewed elsewhere (Farrugia *et al.*, 2005; Dolan, 2006; Ironside, 2006 and Clarke *et al.*, 2007).

To date, a total of 203 probable, or definite, cases of vCJD have been reported worldwide, of which 166 have arisen in the UK, 23 in France, four in Eire and Spain, three in the USA, and one in each of Holland, Portugal, Italy, Saudi Arabia, Japan and Canada (<http://www.cjd.ed.ac.uk/vcjd/world.htm>).

**Correspondence:** Prof. C. A. Ludlam, Department of Haematology, Royal Infirmary, Little France Crescent, Edinburgh EH16 4SA, UK. E-mail: Christopher.Ludlam@ed.ac.uk

First published online 20 October 2008  
doi:10.1111/j.1365-2141.2008.07376.x

Journal Compilation © 2008 Blackwell Publishing Ltd, *British Journal of Haematology*, 123, 18-23

Of these, two of the first and largest outbreaks in the UK and Japan are thought to have resulted from contaminated third US case is thought to have resulted from contaminated plasma. The other cases are thought to have resulted from either countries of origin either through contaminated donor or exported animal or animal products. The current vCJD appears to have reached a plateau in the UK. The risk has waned such that in 2005, the estimated number of cases through the frequency of new cases was estimated to be in France and Spain. All clinically significant vCJD cases have been methionin homozygous at the polymorphic prion protein gene (*PRNP*). Mathematical models suggest that the current incidence of vCJD suggests that there may be an estimate of 70 further cases (39% of those who are homozygous) (Clarke & Urban, 2003). This review also considers the estimate, however, if individuals who are heterozygous are also capable of being infected, the number of transmissions occur from asymptomatic individuals.

Two observations give pause for thought. First, the median age of onset of clinical disease has not been altered over the past 10 years as expected if the number of individuals were exposed to infection over the course of time. The best mathematical model for age-related exposure to infection is that the number of individuals second to the data have a bimodal distribution (see appendix). Miller *et al.* (2007) have shown that the maximum likelihood estimate of the age of onset of clinical infection is not significantly different from that around 50% of heterozygous individuals are homozygous pre- or sub-clinical infection. This is consistent with experimental studies in mice and studies in patients with hereditary prion disease which suggest that individuals who are heterozygous are homozygous at random. 19 have been reported and a lower incidence of development of disease in those who are heterozygous. Miller *et al.* (2007) have observed that give rise to concern that the current cohort of individuals, may be a non-representative population in the UK, may have a higher proportion of

and so a risk of transmitting the disease through blood and tissue products or surgical and medical instrumentation, despite being aseptic invasive.

As there is no currently accepted blood test that reliably identifies vCJD-infected individuals (see below), further studies have been carried out to try to refine the estimate of the prevalence of sub-clinical disease. The National Anonymised Tonsil Archive aims to test 100 000 tonsil samples. Currently, there have been no confirmed positive samples out of 45 000 tested ([http://www.hpa.org.uk/infections/topics\\_az/cjd/tonsil\\_archive.htm](http://www.hpa.org.uk/infections/topics_az/cjd/tonsil_archive.htm)). However, there are reservations around the interpretation of these data, given that the sensitivity of the assays in detecting subclinical vCJD is uncertain, the frequency of involvement of the tonsil as a site of pre-clinical infection is unknown, and a large proportion of the study population are too young to have been exposed to dietary bovine spongiform encephalopathy (BSE). The Spongiform Encephalopathy Advisory Committee (SEAC) has therefore not felt it appropriate to amend the current prevalence estimates within the UK at present (<http://www.seac.gov.uk>).

#### Infectivity in the peripheral blood

Infectivity remains undetectable in the peripheral blood of patients with vCJD despite the fact that clinical transmission has clearly occurred. This apparent contradiction is probably explained by the presence of a species barrier between man and mouse and the limited volumes of blood that can be inoculated in these animals.

Studies in hamsters infected with the 263K strain of scrapie showed similar results to those in the Fukuoka-1 GSS strain in mice (Peden *et al.*, 1998; Ludlam & Turner, 2005), with a point estimate of 1–10 infectious doses (ID<sub>50</sub>) of whole blood of which around 40% was associated with the leucocytes and most of the remainder in the plasma (Gorgoni *et al.*, 2004). Further studies in this model suggest that the majority of cell-associated infectivity is only loosely bound and can be washed off and therefore that the plasma form of infectivity probably predominates. Further studies in mice suggest that the level of infectivity is similar in vCJD-infected animals (Cervenakova *et al.*, 2004). Studies in sheep naturally infected with scrapie, or experimentally infected with BSE, suggest a transmission frequency of up to 50% from blood taken during the preclinical or clinical phase of disease and transfused into recipients from a scrapie-free flock (Hunter *et al.*, 2002). BSE has also been transmitted through butyric acid to the primate *Macaca mulatta* (Boms *et al.*, 2002).

#### Variant CJD transmission by blood transfusion

Within the UK, the Transfusion Medicine Epidemiology Review (TMER) has proved an effective system for collating evidence of possible transmission of vCJD by blood components (Howitt *et al.*, 2006). The UK CJD Surveillance Unit in

Edinburgh shares information about new cases of vCJD with the Blood Transfusion Services, which search their databases to ascertain whether these patients have been blood donors in the past. In this event attempts are made to identify the fate of the blood components (<http://www.cjd.ed.ac.uk/TMER>) and trace, notify and monitor living recipients. The 'reverse' arm of the TMER study attempts to identify which individuals who develop vCJD have received blood transfusions and to identify the donors.

Eighteen patients with vCJD have, or had previously, been blood donors, from whom a total of 66 recipients have been identified, 26 of whom are still alive. Of those who have died, four cases of transmission of vCJD prions have been identified (see below). Many of these patients however will have died of their underlying conditions within 5 years of the implicated transfusion and will not have had time to show clinical evidence of vCJD if infected.

The first symptomatic case of vCJD disease associated with blood transfusion was identified in December 2003. This individual developed vCJD 6.5 years after transfusion of red cells donated by an individual who developed symptoms of vCJD 3.5 years after donation (Llewelyn *et al.*, 2004).

A second case of transmission was identified a few months later in a recipient of red cells from a donor who developed symptoms of vCJD 18 months after donation. This patient died from causes unrelated to vCJD 5 years after transfusion. Postmortem investigations found abnormal prion protein accumulation in the spleen and a cervical lymph node, but not in the brain, and no pathological features of vCJD were found (Peden *et al.*, 2004).

A third patient developed symptoms of vCJD 6 years and died 8.7 years after receiving a transfusion of red blood cells from a donor who developed vCJD about 20 months after this blood was donated (Health Protection Agency 2006).

The fourth case of transmission developed symptoms of vCJD 8.5 years after receiving a transfusion of red blood cells from a donor who developed vCJD about 17 months after this blood was donated. The donor to this patient also donated the vCJD-implicated blood transfused to the third patient (Editorial Team, 2007).

All four patients received transfusions of non-leucodepleted red blood cells between 1996 and 1999. Since October 1999, leucocytes have been removed from all blood used for transfusion in the UK.

These data therefore demonstrate clearly that non-leucodepleted red cells from asymptomatic individuals incubating vCJD can transmit the infection by blood transfusion to other individuals and that the risk of them doing so is relatively high.

#### Donor deferral criteria

There has been little substantive change in blood donor criteria since our previous review (Ludlam & Turner, 2005). Whilst other countries continue to defer those who have spent more than a specified cumulative period of time in the UK, within

the UK only those considered by the CJD Incidents Panel to be 'at risk of vCJD for public health purposes' on account of exposure to implicated surgical instruments, blood components or plasma products, and those who themselves have received blood components, are deferred ([http://www.hpa.org.uk/infections/topics\\_az/cjd/](http://www.hpa.org.uk/infections/topics_az/cjd/)). There is considerable complexity relating to the introduction of similar donor deferral criteria in the context of cell, tissue and organ donation. Broadly, whilst all forms of donation are excluded for patients with CJD or those considered potentially infected, donation of haematopoietic stem cells and solid organs is permitted from those considered 'at risk for public health purposes' and those previously transfused, subject to a risk assessment that weighs the risk of vCJD transmission against the potentially life-saving nature of an otherwise suitable transplant. Donation of other tissues is based on the same donor deferral criteria as blood. Donor deferral criteria remain, however, blunt risk management tools with potential deleterious effects on blood, tissue and organ supply.

#### Importation of blood components

Since our last report (Ludlam & Turner, 2005), the use of imported methylene-blue treated fresh frozen plasma (FFP) has been extended to all patients under the age of 16 years and to high users. Solvent detergent-treated FFP is recommended for patients undergoing plasma exchange for thrombotic thrombocytopenic purpura on the grounds that there is some evidence to suggest that methylene-blue treated FFP has a deleterious impact on outcome in this patient group (Alvarez-Larran *et al.*, 2004). Consideration continues to be given around the possibility of importing FFP and cryoprecipitate for additional groups of patients. Importation of platelets is likely to be impractical given the short shelf-life of these products. However, it may be possible to import red cell concentrates for some groups of patients, for example for children up to 16 years of age. Consideration also has to be given to cost, quality and regulatory requirements and countervailing risks of transmission of other infectious diseases or of component shortages.

#### Advances in the development of a screening test

As previously noted (Ludlam & Turner, 2005), neither nucleic acid transmission nor immunological responses have been clearly identified in association with transmission of prion diseases, rendering standard molecular and serological screening assays unfeasible. Surrogate markers, such as 14-3-3, S100 and erythroid differentiation-related factor, have thus far proved insufficiently sensitive and specific to be of clinical value. Considerable progress has however been made in the development of assays for the abnormal conformer of prion protein, PrP<sup>Sc</sup>.

Normal prion protein (PrP<sup>C</sup>) is a widely expressed 35 kDa 230 amino acid glycosyl-phosphatidylinositol anchored mem-

brane protein. PrP<sup>Sc</sup> is a highly stable, beta-sheet rich, 33 kDa protein that is resistant to proteolysis and is highly infectious. PrP<sup>Sc</sup> is produced from PrP<sup>C</sup> by a process of autocatalytic conversion. PrP<sup>Sc</sup> is highly infectious and is transmitted by ingestion of contaminated meat, direct contact with infected individuals, and by iatrogenic means. PrP<sup>Sc</sup> is highly resistant to heat, formalin, and autoclaving. PrP<sup>Sc</sup> is highly infectious and is transmitted by ingestion of contaminated meat, direct contact with infected individuals, and by iatrogenic means. PrP<sup>Sc</sup> is highly resistant to heat, formalin, and autoclaving. PrP<sup>Sc</sup> is highly infectious and is transmitted by ingestion of contaminated meat, direct contact with infected individuals, and by iatrogenic means. PrP<sup>Sc</sup> is highly resistant to heat, formalin, and autoclaving.

Immunocapillary electrophoresis was amongst the first methods that claimed to be able to detect PrP<sup>TSE</sup> in the peripheral blood. The test material is treated with proteinase and subject to a competitive antibody inhibition assay using a labelled peptide (as the competitor) and a monoclonal antibody that recognises both PrP<sup>TSE</sup> and the peptide (Schmerr *et al.*, 1999; Yang *et al.*, 2005). The technique has however proved difficult to reproduce in other laboratories and failed to discriminate between infected and uninfected blood samples in a blinded study (Cervenakova *et al.*, 2003b).

**Epitope unmasking/masking.** More success has been achieved with the conformation-dependent immunoassay (CDI), which is predicated on the observation that some PrP epitopes are masked within the PrP<sup>TSE</sup> aggregate. An increase in signal intensity produced by a labelled monoclonal antibody by a sample denatured using guanidine hydrochloride when compared with the native (un-denatured) sample denotes the presence of PrP<sup>TSE</sup> (PrP<sup>C</sup> gives the same signal intensity under both conditions). The sensitivity of the technique is increased through the use of highly sensitive dissociation-enhanced lanthanide fluorescence immunoassay for antibody detection and, in some versions of the assay, the use of PK to reduce background signal (Safar *et al.*, 1998, 2002). CDI appears to achieve greater sensitivity than immunoblot (Bellon *et al.*, 2003) and, in the format including PK, may approximate the sensitivity of infectivity assays (Bruce *et al.*, 2001). In the absence of PK it appears able to detect PK-sensitive forms of PrP<sup>TSE</sup>, though it remains unclear as to whether these are infectious or not (Bellon *et al.*, 2003).

The epitope-protection assay developed by Amorfis uses a chemical modification process which alters epitopes on normal PrP but not those buried within PrP<sup>TSE</sup> aggregates. The latter are then disaggregated and the conserved epitopes detected using immunodetection methods (<http://www.amorfis.com>).

PeopleBio have developed an approach where a single antibody is used for both capture and detection steps leading to the blocking of available epitopes by the capture of PrP<sup>C</sup> but not PrP<sup>TSE</sup>.

**PrP<sup>TSE</sup>-specific monoclonal antibodies.** Several antibodies have now been developed that appear to be specific for conformation-dependent epitopes present in PrP<sup>TSE</sup> but not PrP<sup>C</sup> (Korth *et al.*, 1997; Paramithiots *et al.*, 2003; Curin Serbec *et al.*, 2004; Zou *et al.*, 2004). On these, the antibody 15B3, described by Korth *et al.* (1997) and manufactured by Prionics, is the best characterised and has proved capable of detecting infectivity in the peripheral blood of scrapie-infected sheep and BSE-infected cattle in the absence of PK digestion ([http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1\\_9.ppt](http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1_9.ppt)). Three other antibodies (Paramithiots *et al.*, 2003; Curin Serbec *et al.*, 2004; Zou *et al.*, 2004) also appear specific to PrP<sup>TSE</sup> but have not yet been translated to routine assay format.

**PrP<sup>TSE</sup>-specific ligands.** A variety of other ligands have been shown to bind selectively to the abnormally conformed molecule. Plasminogen has been proposed as a means of selective binding PrP<sup>TSE</sup>, but as it can also bind to a variety of other proteins it is therefore unlikely to be sufficiently specific for assay development (Fischer *et al.*, 2000).

Polyanionic compounds are known to selectively bind PrP<sup>TSE</sup> and this property has been employed in the Seprion assay (Lane *et al.*, 2003), which uses coated magnetic beads to capture the molecule. The assay is not dependent on PK treatment and is not species-specific provided a suitable detection antibody is used. It is licensed for postmortem diagnosis of BSE and Chronic Wasting Disease and is reported to be able to distinguish between infected and uninfected blood in scrapie-infected sheep and a small number of human samples.

The approach developed by BioMerieux involves PK digestion, precipitation and denaturation followed by reticulation by streptomycin, chemical capture by calyx-6-arene and detection of the macromolecular aggregates by labelled monoclonal antibody ([http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1\\_9.ppt](http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1_9.ppt)). Detection of PrP<sup>TSE</sup> in a small number of plasma samples from scrapie-infected sheep, BSE-infected cattle and CJD-infected humans has been reported.

Adlyfe have developed a third approach utilising a synthetic peptide based on the region of the PrP molecule involved in the PrP<sup>C</sup>-PrP<sup>TSE</sup> conformational transition. The peptide sequence is coupled to its mirror image as a palindromic molecule fluorescently labelled at each end. When incorporated into PrP<sup>TSE</sup> the peptide folds into a hairpin with a beta-sheet conformation and the fluorophores stack and change their fluorescence wavelength. Further, the folded ligand induces further molecules to adopt the folded conformation and thus amplifies the signal (Grosset *et al.*, 2005). The assay is reported to have discriminated infected from uninfected plasma in natural and experimental scrapie, BSE and CJD.

Chiron have utilised ([http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1\\_9.ppt](http://www.fda.gov/ohrms/dockets/AC/06/slides/2006-4240S1_9.ppt)) a synthetic PrP polypeptide to capture PrP<sup>TSE</sup> on magnetic beads with detection by monoclonal antibody in an ELISA format.

**Amplification.** Two methods have been used to amplify the detection signal. Screening for intensively fluorescent targets utilises double labelled antibodies, more of which bind to PrP<sup>TSE</sup> aggregates than to PrP<sup>C</sup> and giving rise to a stronger fluorescence signal (Bieschke *et al.*, 2000). Immuno-polymerase chain reaction (PCR) also provides a method of amplifying the signal from an antibody or ligand conjugated to a nucleotide sequence utilising the PCR (Barletta *et al.*, 2005).

Two further approaches have been developed that result in the amplification of PrP<sup>TSE</sup> itself. The first of these, protein misfolding cyclic amplification (PMCA) has given rise to considerable excitement. PrP<sup>TSE</sup> seeded into an excess of PrP<sup>C</sup> leads to formation of new PrP<sup>TSE</sup>. That PrP<sup>TSE</sup> is then

fragmented through sonication or shaking and leads to a new round of PrP<sup>TSE</sup> formation (Kocisko *et al.*, 1994; Saborio *et al.*, 2001). Recurrent cycles therefore of incubation and fragmentation lead to amplification of the original PrP<sup>TSE</sup> (Castilla *et al.*, 2005). Immunoblot and CDI have been used for detection of PrP<sup>TSE</sup> and infectivity. Studies show that 146 sonication cycles produced an increase in signal intensity of around 6000-fold, whilst a second 'nested' set of 118 cycles with a fresh source of normal PrP led to an approximate 10<sup>7</sup> fold amplification. The technique has proved capable of discriminating infected from uninfected blood from hamsters experimentally infected with scrapie, however there are recent reports of detection of PrP<sup>TSE</sup> in uninfected animal brain implying the possibility of low levels of abnormally conformed PrP in 'normal' individuals.

A number of cell-based amplification techniques have been described in which the rodent cell lines N2a (Nishida *et al.*, 2000), PK-1 (Klohn *et al.*, 2003), Rov-9 (Birkett *et al.*, 2001) and CAD-5 are infectable by natural or experimental strains of scrapie and demonstrate amplification of PrP<sup>TSE</sup> detected by immunoblot. No cell-based amplification has yet been successfully reported for CJD.

Both these kinds of amplification take several days (PMCA) to weeks (cell-based assays) and would therefore be better positioned as confirmatory rather than screening assays.

**Considerations with regard to assay assessment.** Whilst the above is not a comprehensive list of all the assays under development, it does provide a flavour of the range and variety of approaches and their relative strengths and weaknesses. Some of these are now approaching the point at which they may be Council of Europe (CE) marked and marketed as potential clinical assays. There are, therefore, a series of further considerations relating to the potential assessment and utility of prion assays prior to clinical implementation.

The required sensitivity is difficult to gauge because the level, spatial distribution and temporal variation of infectivity in the blood of patients with vCJD or healthy individuals with subclinical infection is unknown. The generalizability of experimental data from mouse and hamster experiments to the human condition cannot be assumed (Castilla *et al.*, 2005). Moreover, the relationship between infectivity and PrP<sup>TSE</sup> is complex. Although many authorities believe PrP<sup>TSE</sup> to be

central, there is evidence that it may be present at very low levels in the peripheral blood of individuals who are not yet clinically affected (e.g. 10<sup>-10</sup> to 10<sup>-12</sup> g/l range) (Saborio *et al.*, 2001). The latter is in agreement with the low infectivity of blood from such individuals (10<sup>-10</sup> to 10<sup>-12</sup> ID<sub>50</sub>/ml) (Saborio *et al.*, 2001). It is reasonable to assume that the infectious material in such individuals is not detectable by the current methods of detection of vCJD in peripheral blood from natural animals or a small number of patients with vCJD in peripheral blood from natural animals (Birkett *et al.*, 2001) and that the sensitivity of prion assays (Birkett *et al.*, 2001).

Where the sensitivity of a technical assay exceeds the specific population into which it is likely to be highly specific (i.e. a high positive predictive value), the sensitivity of prion assays (Birkett *et al.*, 2001) can be very poor, giving rise to the number of false positives being much greater than the number of true positives (Fig. 1).

The 'Highly Sensitive' assay Facility, containing several different assays, is currently being developed and evaluated in the general population. Finally, there are a number of things to be considered in relation to subclinical infection. Such an assay would need to have a very high sensitivity to detect the presence of subclinical infection. However, it is difficult to gauge because the level, spatial distribution and temporal variation of infectivity in the blood of patients with vCJD or healthy individuals with subclinical infection is unknown. The generalizability of experimental data from mouse and hamster experiments to the human condition cannot be assumed (Castilla *et al.*, 2005). Moreover, the relationship between infectivity and PrP<sup>TSE</sup> is complex. Although many authorities believe PrP<sup>TSE</sup> to be

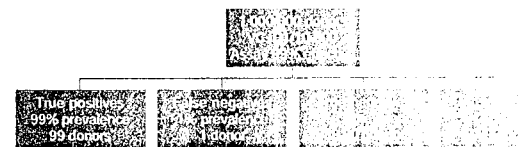


Fig. 1. Outcome of screening of a 'normal' population of one million persons to determine the presence of a variant Creutzfeldt-Jakob disease using an assay with a sensitivity and specificity of 99%. Although the number of false positives greatly exceeds the number of true positives.

blood donors would not be able to continue to donate if they have a positive result. It is likely, for example, to take the donation with this result as a medical investigation, even if the donor consents to such a strategy. Test-positive individuals will therefore have to be told of the outcome and (presumably) managed as 'at risk for public health purposes'. Clearly this will cause significant distress and an increase in psychological and social problems for some people, act as a disincentive to blood donation and therefore a negative impact on the blood supply. Moreover, it is likely that previous recipients of blood components from these donors will also have to be traced and contacted (if possible), giving rise to a much larger group of individuals in the population considered 'at risk of public health purposes' and requiring specific precautionary measures to be taken in the event of a future medical investigation (see below). A comprehensive health and economic evaluation will therefore have to weigh the positive impact of reducing potential secondary transmission of vCJD against these potential negative consequences.

#### Blood component processing

When leucodepletion was introduced in the UK in 1999 as a means to reduce the risk of secondary transmission of vCJD, UK confidential data from mice infected with the PrP<sup>Sc</sup> agent of scrapie (Mead & Straüssler-Scheinker disease) (Hewitt *et al.*, 2001) suggests that leucodepletion filters have little impact on plasma-borne infectivity. Studies in the 2008 (Lambert *et al.*, 2004) similarly suggest a 40–70% reduction in whole blood infectivity, consistent with the removal of leucocyte-associated infectivity, but not that present in the plasma. Table 1 illustrates the likely distribution of residual infectivity in a unit of leucodepleted red cell concentrate prepared by bottom and top processing method (with a residual plasma volume of around 10–15 ml). Assuming 10 ID/ml infectivity in whole blood, just over 130 ID would be left in the unit and that up to a 3 log further reduction is required to impact upon the risk of transmission (i.e. achieve <1 ID/unit). Red cell concentrates prepared by the more common top-top methodology contain greater amounts of residual plasma (around 20 ml) and would consequently require a 4-log reduction. The absence of data on the level of infection in human blood means an uncertainty of at least 1-log at and above point estimates. It can be said in summary, however, that it is unlikely that current blood component processing will suffice to reduce the risk of transmission in most plausible infectivity scenarios.

Three companies are working on the development of prion reduction filters. One has a CE-marked dCp-on filter which is used in series with a leucodepletion filter. Published studies using this filter material show a 3-log reduction in infectivity on brain homogenate spikes and to the limit of detection (0.1 ng) in endogenous infectivity studies (Gregori *et al.*, 2004). Two other companies are working on the development of combined leucodepletion/prion reduction filters. All prion

**Table 1.** Residual infectivity distribution in a unit of leucodepleted red cell concentrate.

Log reduction in infectivity	Residual leucocytes	Residual plasma	Residual infectivity
Leucodepletion alone	0.2	130	130.2
1 Log	0.2	13	13.2
2 Log	0.2	1.3	1.5
3 Log	0.2	0.13	0.33
4 Log	0.2	0.013	0.213

The data represents the likely distribution of residual infectivity in a unit of leucodepleted red cell concentrate prepared by a bottom and top processing method (with a residual plasma volume of around 10 ml).

Assuming 10 ID/ml infectivity in whole blood with 40% (i.e. 4 ID/ml) being removed by leucodepletion and the remainder residing in the plasma (i.e. for a haematocrit of 0.45 a plasma concentration of approximately 13 ID/ml), around 130 ID remains in the unit's plasma. Hence up to approximately a 3 log further reduction is required to reduce the risk of transmission to <1 ID/unit.

reduction filters will have to undergo independent assessment of clinical safety and efficacy within a series of studies managed by the UK and Irish Blood Services and agreed with SEAC and the Advisory Committee on the Safety of Blood, Tissues and Organs (<http://www.advisorybodies.doh.gov.uk/acsbto/index.htm>). Part of the problem for both manufacturers and Blood Services is the absence of assays capable of detecting either PrP<sup>Sc</sup> or infectivity in the peripheral blood of patients with vCJD. Assessment of the efficacy of the technology is therefore based on brain homogenate spikes (where baseline infectivity is sufficient to detect a 3–4 log reduction but the physico-chemical form of the spike is unlikely to be similar to that of plasma based infectivity), and endogenous infectivity studies (where the form of infectivity is likely to be more relevant, but the baseline infectivity is sufficiently low that little more than a 1-log reduction is detectable). There remain, therefore, fundamental questions relating to the clinical relevance of different forms of spike material and general applicability of these kinds of studies to the human situation. The potential for deleterious effects on the red cell concentrate itself are also a matter for concern, both in terms of the possibility of alterations to the rheological or antigenic profile of the red cells and the loss in the volume of the additional filter. The latter would have a particular impact if used in conjunction with bottom and top processing, the combined effect of which may reduce the red cell mass in a concentrate below current standards, necessitating additional transfusions for some individuals.

With regard to platelet concentrates, re-suspension in optimal additive solution rather than plasma would reduce the amount of residual plasma by around 65% to 80–90 ml. This would still contain more than enough infectivity to transmit infection to the recipient under even the most

optimistic of the current infectivity assumptions and is likely to be ineffectual. Prion reduction filters are also currently applicable to either platelet concentrates or FFP.

#### Plasma product manufacturing

It is reassuring that to date no recipient of a pooled plasma product has developed vCJD. However in 1997, shortly after the first description of vCJD as a new condition, there was concern that the UK plasma supply might have the potential to transmit the infectious agent and that plasma collected from countries where there were few or no cases of vCJD might pose a lower risk (Ludlam, 1997). Although this view gave rise to controversy, the regulatory authorities moved to a position of allowing, and subsequently mandating that pooled plasma products manufactured in the UK should only be made from plasma imported from parts of the world at low risk of vCJD.

In an attempt to help define the risk of PrP<sup>Sc</sup> transmission by plasma-derived products, detailed studies have been undertaken to assess how prions are partitioned during the plasma fractionation process, mainly by spiking the starting plasma with 'exogenous' prion derived from brain homogenates of experimentally infected animals. The strengths and weaknesses of this approach are similar to those described above in the discussion around the assessment of prion filters. In general there was least clearance of prion in the manufacture of factor VIII, IX and antithrombin concentrates, greater clearance in the preparation of intravenous immunoglobulin and greatest clearance in the manufacture of albumin (Hewitt, 1999).

The way in which different countries responded to the risk that plasma products might transmit the infectious agent varied and depended partly on the perceived relative number of donors who might be infectious as well as details of the plasma fractionation techniques used in each country.

In the UK, using data on partitioning of prion infectivity during manufacture of plasma products, along with the animal data on the likely range of infectivity in individuals with sub-clinical infection, a risk assessment was undertaken to quantify the risk of recipients of such products being infected. The CJD Incidents Panel have taken the view that an individual with a >1% additional risk of exposure to an infectious dose of vCJD should be notified and managed as 'at risk for public health purposes'.

To date a total of 174 'implicated' batches of plasma products have been identified as having been manufactured from a pool of plasma to which an individual contributed who subsequently developed vCJD (Hewitt *et al.*, 2006). For each of these batches a detailed risk assessment was carried out that included the total number of donations included in the pool, the details of the plasma fractionation process used during manufacture and (conservative) estimates of the likely cumulative reduction in infectivity over the manufacturing process. The outcome was expressed as the likely mass of product to which an individual would have had to be exposed to increase

their risk of exposure to an infectious dose of vCJD to a level of 1% above their baseline risk. The 'implicated' batches were identified by tracing back the manufacturing process of each product to the donor who had developed vCJD, where this was possible, and then tracing back the manufacturing process of each product to the donor who had contributed to the pool of plasma from which the product was manufactured. The 'implicated' batches were those for which the donor who had developed vCJD had contributed to the pool of plasma from which the product was manufactured. The 'implicated' batches were those for which the donor who had developed vCJD had contributed to the pool of plasma from which the product was manufactured. The 'implicated' batches were those for which the donor who had developed vCJD had contributed to the pool of plasma from which the product was manufactured.

Confidentiality was maintained throughout the process for public health purposes. The specific details of the manufacturing process, including the names of the individuals who had contributed to the pool of plasma from which the product was manufactured, were agreed to be kept confidential. The specific details of the manufacturing process, including the names of the individuals who had contributed to the pool of plasma from which the product was manufactured, were agreed to be kept confidential. The specific details of the manufacturing process, including the names of the individuals who had contributed to the pool of plasma from which the product was manufactured, were agreed to be kept confidential. The specific details of the manufacturing process, including the names of the individuals who had contributed to the pool of plasma from which the product was manufactured, were agreed to be kept confidential.

Within the UK, all implicated batches of plasma products, surgical, medicinal and transfusion, were notified to the Advisory Committee on the Safety of Blood, Tissues and Organs. The Incidents Panel, Institute of Blood Transfusion, where there were no cases of vCJD, were notified of the implicated batches and the system and information used to identify the implicated batches. The implicated batches were notified to the Advisory Committee on the Safety of Blood, Tissues and Organs. The implicated batches were notified to the Advisory Committee on the Safety of Blood, Tissues and Organs. The implicated batches were notified to the Advisory Committee on the Safety of Blood, Tissues and Organs.

difficulties in the performance of biopsies with gastrointestinal endoscopes because the samples obtained would probably contain lymphoid tissue. The financial implications are significant because the endoscopes cannot be decontaminated and must effectively be discarded. Both upper and lower gastrointestinal endoscopies without biopsy do not result in the instrument being considered as potentially 'contaminated' and it can therefore be reused on other patients after standard cleaning procedure. The concern about possible contamination of instruments has also led to an increased use of capsule endoscopies, which give good images but cannot be used to biopsy or treat gut lesions.

Although no individuals with haemophilia have thus far developed vCJD and a retrospective study of autopsy samples from individuals with haemophilia in 1998 showed no evidence of sub-clinical infection, it has been important to try and gather more data (Lee *et al.*, 1998). This has not been easy and depends upon procuring appropriate tissue samples prospectively from individuals undergoing clinically necessary surgery in addition to consent for autopsy. In addition it has been important to try and develop a record of the extent of exposure of individuals to 'implicated' batches of concentrate, as well as all recipients of UK clotting factor concentrates over the 22-year period of exposure. This is being co-ordinated by UK Haemophilia Centre Doctors' Organisation by accumulating the data for subsequent anonymised studies.

#### Communication with patients and the general public

Keeping recipients of blood and blood products informed about the current state of knowledge and in particular informing individuals about their individual risks has proved challenging because of the complexity and uncertainty inherent in our understanding of the field. It has been important for there to be close collaboration between those able to assess the risk of vCJD infection, physicians responsible for clinical services and patient organisations representing those potentially affected. For those who have received blood components from donors who subsequently developed vCJD, the risk of exposure to vCJD is judged to be high and these individuals have been contacted on an individual basis and offered counselling and specialist follow-up. Similarly, blood donors who have donated blood administered to a patient who later developed vCJD have been contacted and are managed as 'at risk for public health purposes'. In 2004, all patients with haemophilia were sent a letter stating whether or not they had or had not received UK plasma-derived clotting concentrates between 1980 and 2001, irrespective of whether or not they had received UK plasma products, because in an earlier mailing about this topic only those in the 'at risk' group were contacted and this left non-recipients of letters not knowing whether they had not been potentially exposed or whether their letter had got lost in the post. All were offered the opportunity for individual counselling. It is this attention to the detail of how

patients are informed that is critical in trying to ensure that individuals feel confident in the arrangements.

For patients potentially exposed to other implicated plasma products, the issue of traceability and notification have proved more problematic. Whilst patients with primary immunodeficiency share a similar close long-term relationship with their physicians, those receiving immunoglobulin for other clinical indications or high doses of albumin (for example during plasma exchange), are often discharged following their acute care. The absence of a general system of traceability for plasma products and of searchable clinical notes has made the follow-up of the latter groups of potentially exposed patients highly problematic.

#### Concluding remarks

Three years after our last review (Ludlam & Turner, 2005), the management of the risk of transmission of vCJD by blood and plasma products remains highly challenging. Whilst the diminishing number of clinical cases is reassuring, there are continuing uncertainties surrounding the prevalence of sub-clinical disease, the level of infectivity in peripheral blood of such individuals, and the overall risk of transmission and development of clinical disease. Much progress has been made in the development of new technologies, such as prion filters and prion assays, but assessment of these is problematic and cost and countervailing risks need to be considered. Accurate and timely communication with the general public and with those who are considered to be at increased risk of exposure remains essential given the continuing complexity and uncertainty of the field.

#### References

- Alvarez-Larran, A., Del Rio, J., Ramirez, C., Albo, C., Peña, F., Campos, A., Cid, J., Muncunill, J., Sastre, J.L., Sanz, C. & Pereira, A. (2004) Methylene blue-photoinactivated plasma vs. fresh-frozen plasma as replacement fluid for plasma exchange in thrombotic thrombocytopenic purpura. *Vox Sanguinis*, **86**, 246–251.
- Barietta, J.M., Edelman, D.C., Highsmith, W.E. & Constantine, N.T. (2005) Detection of ultra-low levels of pathological prion protein in scrapie infected hamster brain homogenates using real-time immuno-PCR. *Journal of Virological Methods*, **127**, 154–164.
- Bellon, A., Seyfort-Brandt, W., Lang, H., Baron, H., Groner, A. & Vey, M. (2003) Improved conformation dependent immunoassay: suitability for enhance prion detection with enhanced sensitivity. *The Journal of General Virology*, **84**, 1921–1925.
- Bieschke, J., Giese, A., Schulz-Schaeffer, W., Zerr, I., Poser, S., Eigen, M., Eigen, M. & Kretzschmar, H. (2000) Ultrasensitive detection of pathological prion protein aggregates by dual colour scanning for intensely fluorescent targets. *Proceedings of the National Academy of Sciences of the United States of America*, **97**, 5468–5473.
- Birkett, C.R., Hennion, R.M., Bembridge, D.A., Clarke, M.C., Chree, A., Bruce, M.E. & Bostock, C.J. (2001) Scrapie strains maintain biological phenotypes on propagation in a cell line in culture. *EMBO Journal*, **20**, 3351–3358.

- Bons, N., Lehmann, S., Mestre-François, N., Dormont, D. & Brown, P. (2002) Brain and buffy coat transmission of bovine spongiform encephalopathy to the primate *Microcebus murinus*. *Transfusion*, **42**, 513–516.
- Brown, P., Rohwer, R.G., Dunstan, B.C., MacAuley, C., Gajdusek, D.D. & Drohan, W.N. (1998) The distribution of infectivity in blood components and plasma derivatives in experimental models of transmissible spongiform encephalopathy. *Transfusion*, **38**, 210–217.
- Brown, P., Cervenkova, L., McShane, L.M., Darcey, P., Rubenstein, J. & Drohan, W.N. (1999) Further studies of blood infectivity in an experimental model of transmissible spongiform encephalopathy with an explanation of why blood products do not transmit Creutzfeldt-Jakob disease in humans. *Transfusion*, **39**, 1169–1178.
- Bruce, M.E., McConnell, I., Will, R.G. & Ironside, J.W. (2001) Detection of variant Creutzfeldt-Jakob disease infectivity in extra-neural tissues. *Lancet*, **358**, 208–209.
- Castilla, J., Saa, P., Hetz, C. & Soto, C. (2005) In vitro generation of infectious scrapie prions. *Cell*, **121**, 195–206.
- Castilla, J., Saa, P., Morales, R., Abid, K., Maundrell, K. & Soto, C. (2006) Protein misfolding cyclic amplification for diagnosis and prion propagation studies. *Methods in Enzymology*, **412**, 3–21.
- Cervenkova, L., Yakovleva, O., McKenzie, C., Koldchinsky, S., McShane, L., Drohan, W.N. & Brown, P. (2003a) Similar levels of infectivity in the blood of mice infected with human-derived vCJD and GSS strains of transmissible spongiform encephalopathy. *Transfusion*, **43**, 1687–1694.
- Cervenkova, L., Brown, P., Soukhariev, S., Yakovleva, O., Dirlikov, H., Saenko, E.L. & Drohan, W.N. (2003b) Failure of immunocompetitive capillary electrophoresis assay to detect disease specific prion protein in buffy coat from humans and chimpanzees with Creutzfeldt-Jakob disease. *Electrophoresis*, **24**, 853–859.
- Clarke, P. & Ghani, A.C. (2005) Projections of the future course of the primary vCJD epidemic in the UK: inclusion of subclinical infections and the possibility of wider genetic susceptibility. *Journal of the Royal Society, Interface*, **2**, 19–31.
- Clarke, P., Will, R.G. & Ghani, A.C. (2007) Is there the potential for an epidemic of variant Creutzfeldt-Jakob disease via blood transfusion in the UK? *Journal of the Royal Society, Interface*, **4**, 675–684.
- Collinge, J., Sidle, K.C.L., Meads, J., Ironside, J. & Hill, A.P. (1996) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **383**, 685–690.
- Curin Seibec, V., Brezjanac, M., Popovic, M., Pretny, Hartman, K., Galvani, V., Ruprecht, R., Cernilec, M., Vranac, T., Hainzer, I. & Jerala, R. (2004) Monoclonal antibody against a peptide of human prion protein discriminates between Creutzfeldt-Jakob's disease affected and normal brain tissue. *Journal of Biological Chemistry*, **279**, 3694–3698.
- Dolan, G. (2006) Clinical implications of emerging pathogens in haemophilia: the variant Creutzfeldt-Jakob experience. *Haemophilia*, **12**(Suppl. 1), 16–20.
- Editorial Team (2007) Fourth case of transfusion-associated variant-CJD. *Euro Surveillance*, **12**, pii. 3117. Available at <http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=3117>.
- Farrugia, A., Ironside, J.W. & Giangrande, P. (2005) Variant Creutzfeldt-Jakob disease transmission by plasma products: assessing and communicating risk in an era of scientific uncertainty. *Vox Sanguinis*, **89**, 186–192.
- Fischer, M.B., Roeckl, C., Parizek, P., Schwarz, H.P. & Aguzzi, A. (2000) Binding of disease-associated prion protein to plasminogen. *Nature*, **408**, 479–488.

- Foster, P. (1997) Use of a monoclonal antibody to detect variant Creutzfeldt-Jakob disease in blood components. *Journal of Virological Methods*, **68**, 229–234.
- Health Protection Agency (2006) Variant Creutzfeldt-Jakob disease: variant CJD. *UK Health Protection Agency, Health Protection Agency, London*, **14**, 1–10.
- Hewitt, P.H., Hannon, G.H., Maclean, J., Clarke, P., Ironside, J.W., Tanshian, M. *Journal of Virology*, **91**, 221–226.
- Hill, A.P., Saika, S., Ghetti, B. & Brown, P. (2000) The prion agent of variant Creutzfeldt-Jakob disease: a highly infectious agent. *Journal of Virology*, **74**, 273–278.
- Hunter, M., Bostock, C., Ghani, A., Hill, A.P., Ironside, J.W., Clarke, P., S. MacKinnon, C., Hannon, G.H. & Brown, P. (2005) Variant Creutzfeldt-Jakob disease by the transfusion route. *Journal of Virology*, **79**, 293–298.
- Ironside, J.W. (2006) Variant Creutzfeldt-Jakob disease: transmission by blood transfusion. *Journal of Virology*, **80**, 143–146.
- Ironside, J.W. & Clarke, P. (2007) A quantitative light scattering assay for the detection of prion protein in plasma. *Journal of Virology*, **81**, 1401–1407.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2004) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2005) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2006) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2007) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2008) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2009) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2010) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2011) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2012) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2013) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2014) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2015) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2016) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2017) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2018) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2019) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2020) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2021) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.
- Ironside, J.W., Brown, P., Hill, A.P., Clarke, P., MacKinnon, C., Hannon, G.H., Tanshian, M., & Ghani, A.C. (2022) Molecular analysis of prion strain variation and the aetiology of new variant CJD. *Nature*, **431**, 85–90.

## Managing the risk of transmission of variant Creutzfeldt-Jakob disease by blood products

Christopher A. Ludlam<sup>1</sup> and Marc L. Turner<sup>2</sup>

<sup>1</sup>Department of Haematology, Royal Infirmary, and <sup>2</sup>South East Scotland Blood Transfusion Centre, Edinburgh, UK

### Summary

Whereas plasma-derived clotting factor concentrates now have a very good safety record for not being infectious for lipid-enveloped viruses, concern has arisen about the possibility that prion diseases might be transmitted by blood products. There is epidemiological evidence that classical sporadic Creutzfeldt-Jakob disease (CJD) is not transmitted by blood transfusion. There is now good evidence that the abnormal prion associated with variant CJD can be transmitted by transfusion of fresh blood components and infect recipients. To reduce the risk of the pathological prion in the UK infecting recipients of clotting factor concentrates, these are now only manufactured from imported plasma collected from countries where there has not been bovine spongiform encephalopathy (BSE) in cattle and the risk of variant CJD in the population is, therefore, considered negligible. The safety of these concentrates is also enhanced because prion protein is, to an appreciable extent, excluded by the manufacturing process from the final product. To help reduce the chance of prion transmission by fresh blood products, donations are leucodepleted, there is increasing use of imported fresh frozen plasma (especially for treating children) and potential donors, who have been recipients of blood since 1980 (the beginning of the BSE epidemic in cattle), are deferred.

**Keywords:** variant Creutzfeldt Jakob disease, transfusion, epidemiology, safety, haemophilia.

Emerging pathogens will always challenge the safety of blood transfusion. Whilst the risk of hepatitis B virus (HBV), hepatitis C virus (HCV) or human immunodeficiency virus (HIV) transmission by blood components and plasma products is now small (<http://www.eurosurveillance.org>), new potentially transfusion-transmissible pathogens continue to emerge.

Many challenges were posed by the emergence of variant Creutzfeldt Jakob disease (vCJD) in 1996 (Will *et al.*, 1996).

Whereas there has been little evidence of sporadic Creutzfeldt Jakob disease (CJD) re-emerging from variant CJD, the transmission of prion protein to a different prion protein (PrP<sup>Sc</sup>) with a wild-type substrate is thought not to occur. It is, therefore, unlikely that classical sporadic CJD (kindred 1978 R. Little *et al.*, 1998) is transmitted to humans via the transfusion of blood components. There is, however, good evidence that the CJD associated with variant CJD (vCJD) can be transmitted by transfusion of fresh blood components to infect recipients. The UK transfusion services have taken steps to reduce the risk of vCJD infecting recipients of clotting factor concentrates. These are now only manufactured from imported plasma collected from countries where there has not been bovine spongiform encephalopathy (BSE) in cattle and the risk of vCJD in the population is, therefore, considered negligible. The safety of these concentrates is also enhanced because prion protein is, to an appreciable extent, excluded by the manufacturing process from the final product. To help reduce the chance of prion transmission by fresh blood products, donations are leucodepleted, there is increasing use of imported fresh frozen plasma (especially for treating children) and potential donors, who have been recipients of blood since 1980 (the beginning of the BSE epidemic in cattle), are deferred.

The transfusion services have also taken steps to reduce the risk of vCJD infecting recipients of clotting factor concentrates. These are now only manufactured from imported plasma collected from countries where there has not been bovine spongiform encephalopathy (BSE) in cattle and the risk of vCJD in the population is, therefore, considered negligible.

The transfusion services have also taken steps to reduce the risk of vCJD infecting recipients of clotting factor concentrates. These are now only manufactured from imported plasma collected from countries where there has not been bovine spongiform encephalopathy (BSE) in cattle and the risk of vCJD in the population is, therefore, considered negligible. The safety of these concentrates is also enhanced because prion protein is, to an appreciable extent, excluded by the manufacturing process from the final product. To help reduce the chance of prion transmission by fresh blood products, donations are leucodepleted, there is increasing use of imported fresh frozen plasma (especially for treating children) and potential donors, who have been recipients of blood since 1980 (the beginning of the BSE epidemic in cattle), are deferred.

Correspondence: Professor Christopher A. Ludlam, Department of Haematology, Royal Infirmary, Little France Crescent, Edinburgh EH16 4SX, UK. E-mail: christopher.ludlam@ed.ac.uk

Ludlam, C.A. & Turner, M.L. (2005) Managing the risk of transmission of variant Creutzfeldt Jakob disease by blood products. *British Journal of Haematology*, **132**, 13–24.

Muir, P., Newham, I., Jones, N., Bergeron, C., Gregori, L., Asher, D., van Engelenburg, P., Stoeckel, T., Vey, M., Barnard, G. & Head, M. (2004) Standards for the assay of Creutzfeldt-Jakob disease specimens. *Journal of General Virology*, **85**, 1777–1784.

Nishida, N., Harris, D.A., Vletter, D., Laude, H., Probst, Y., Grassi, J., Casanova, P., Miklavet, O. & Lehmann, S. (2000) Successful transmission of three mouse adapted scrapie strains to murine neuroblastoma cell lines over expressing wild-type mouse prion protein. *Journal of Virology*, **74**, 320–325.

Paramithiotis, E., Pinard, M., Lawton, T., LeBoissiere, S., Leathers, V.L., Zou, W.Q., Estey, L.A., Lamentagne, I., Lehto, M.T., Kondejewska, I.H., Francoeur, G.P., Papadopoulos, M., Haghghat, A., Spatz, S.J., Hawk, M., White, R.G., Ironside, J., O'Rourke, K., Tonelli, C., Lefebvre, H.L., Casarabian, A. & Castagna, N.R. (2003) A prion protein epitope selective for the pathologically misfolded conformation. *Nature Medicine*, **9**, 893–899.

Pedon, J.H., Frank, M.W., Ritchie, D.B., Bell, J.E. & Ironside, J.W. (2004) Creutzfeldt Jakob after blood transfusion in a PRNP codon 129 heterozygous patient. *Lancet*, **364**, 507–521.

Sabaana, G.P., Heremans, B. & Soto, C. (2001) Sensitive detection of pathological prion protein by cyclic amplification of protein misfolding. *Nature*, **411**, 815–819.

Safar, J., Wille, H., Tre, M., Groth, C., Serban, H., Torchia, M., Cohen, B.E. & Prusiner, S.B. (1998) Eight prion strains have PrP<sup>Sc</sup> molecules with different conformations. *Nature Medicine*, **4**, 1157–1165.

Safar, J.G., Scott, M., Moughan, J., Deering, C., Didorenko, S., Vergara, I., Fall, H., Legnani, G., Jaciere, E., Sforzosi, L., Serban, H., Giam, D., Burton, D.R., Prusiner, S.B. & Williamson, R.A. (2002) Measuring prions causing bovine spongiform encephalopathy or chronic wasting disease by immunoassays and transgenic mice. *Nature Biotechnology*, **20**, 1147–1152.

Safar, J.G., Geschwind, M.D., Deering, C., Didorenko, S., Sattavat, M., Sanchez, H., Serban, A., Vey, M., Baron, H., Giles, K., Miller, B.L., Dearmond, S.J. & Prusiner, S.B. (2005) Diagnosis of human prion disease. *Proceedings of the National Academy of Sciences of the United States of America*, **102**, 3501–3506.

Schmerr, M.J., Jenny, A.L., Bulgin, M.S., Miller, J.M., Hamir, A.N., Cutlip, R.C. & Goodwin, K.R. (1999) Use of capillary electrophoresis and fluorescent labeled peptides to detect the abnormal prion protein in the blood of animals that are infected with a transmissible spongiform encephalopathy. *Journal of Chromatography, A*, **853**, 207–214.

Seitz, R., von Auer, F., Blumel, J., Burger, R., Buschmann, A., Dietz, K., Heiden, M., Hitzler, W.E., Klamm, H., Kreil, T., Kretzschmar, H., Nübling, M., Offergeld, R., Pauli, G., Schottstedt, V., Volkers, P. & Zerr, I. (2007) Impact of vCJD on blood supply. *Biologicals*, **35**, 79–97.

Silveira, J.R., Raymond, G.J., Hughson, A.G., Race, R.E., Sim, V.L., Hayes, S.F. & Caughey, B. (2005) The most infectious prion protein particles. *Nature*, **437**, 257–261.

Wadsworth, J.D., Joiner, S., Hill, A.F., Campbell, T.A., Desbruslais, M., Luthert, P.J. & Collinge, J. (2001) Tissue distribution of protease resistant prion protein in variant Creutzfeldt-Jakob disease using a highly sensitive immunoblotting assay. *Lancet*, **358**, 171–180.

Yang, W.C., Yeung, E.S. & Schmerr, M.J. (2005) Detection of prion protein using a capillary electrophoresis-based competitive immunoassay with laser-induced fluorescence detection and cyclodextrin-aided separation. *Electrophoresis*, **26**, 1751–1759.

Yuan, J., Xiao, X., McGeehan, J., Dong, Z., Cali, I., Fujioka, H., Kong, Q., Kneale, G., Gambetti, P. & Zou, W.Q. (2006) Insoluble aggregates and protease-resistant conformers of prion protein in uninfected human brains. *Journal of Biological Chemistry*, **281**, 34848–34858.

Zou, W.-Q., Zheng, J., Gray, D.M., Gambetti, P. & Shen, S.G. (2004) Antibody to DNA detects scrapie but not normal prion protein. *Proceedings of the National Academy of Sciences of the United States of America*, **101**, 1380–1385.

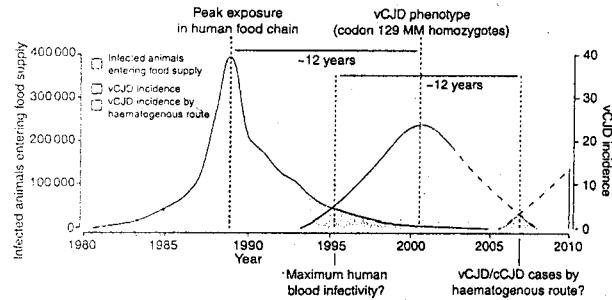


Fig. 1. Incidence of bovine spongiform encephalopathy and variant Creutzfeldt Jakob disease in the UK (---, predicted cases). The right hand peak illustrates the potential for secondary spread by haematogenous route. Reprinted from Collins *et al.* (2004) with permission from Elsevier.

In addition, a critical polymorphism at codon 129 coding for methionine or valine leads to significant variation in the susceptibility to, and incubation period of, human prion diseases. In the UK, 37% of the general population are homozygous for methionine at this locus, 11% are homozygous for valine and 52% heterozygous. Methionine homozygosity is much more common than expected amongst patients with CJD (*vide infra*). PrP is inserted into the cell membrane predominantly via a glycosylphosphatidylinositol (GPI) anchor, although transmembrane and soluble forms have also been described. The glycoprotein is predominantly located in calveolar zones in the cell membrane and is estimated to have a half-life of around 6 h, being internalised into endosomes with a proportion recycling to the cell surface (Shyng *et al.*, 1993). The function of the protein remains unclear, it has been shown to bind to a laminin receptor precursor protein (Martins *et al.*, 1997; Rieger *et al.*, 1997) and act as a copper metalloproteinase (Brown *et al.*, 1997a). PrP null mice appear to develop normally although some strains show subtle neurological abnormalities (Tobler *et al.*, 1996). Prion formation involves changes in the secondary and tertiary conformations of the PrP molecule: up to 40–50% of the molecule can be in the form of beta-pleated sheet, mainly at the expense of the membranedistal unstructured region. This changes the physicochemical properties of the molecule and engenders relative resistance to proteinase digestion. Prion protein aggregates (PrP<sup>Sc</sup>) are deposited in cells and tissues leading to the formation of amyloid-like plaques and in the nervous system to neuronal death, astrogliosis and spongiform change.

The mechanism by which PrP<sup>C</sup> is converted to PrP<sup>Sc</sup> remains unclear, as does its precise role in the aetiology of the disease. The prion hypothesis (Prusiner, 1998) proposes that the PrP<sup>Sc</sup> molecule itself converts PrP<sup>C</sup> to the abnormal conformation, either through a process of heterodimerisation or through nuclear polymerisation (Aguzzi & Weissmann, 1997). PrP<sup>Sc</sup> is relatively resistant to proteinase-K digestion and different molecular strains of disease can be identified by the balance of di-glycosylated, mono-glycosylated and non-glycosylated spe-

cies. Several molecular strains of PrP<sup>Sc</sup> occur in sporadic CJD; however, only a single strain of PrP<sup>Sc</sup> is found in variant CJD, which is similar to that seen in naturally occurring bovine spongiform encephalopathy (BSE) in cattle, and BSE transmitted naturally and experimentally to other animals (Collinge *et al.*, 1996; Hill *et al.*, 1997a). Evidence that variant CJD and BSE represent the same strain of prion disease also stems from infectivity studies in a prion disease strain typing panel of inbred experimental mice, where the patterns of incubation period and neuropathological targeting were similar and differed from those seen in sporadic CJD, scrapie and other prion diseases (Bruce *et al.*, 1997).

### Prion diseases in other species

A range of prion disorders have been described including those involving the Sup35p and Ure2p proteins in yeast, which appear to be non-pathogenic and convey a survival advantage under certain circumstances (Burwinkel *et al.*, 2004).

Scrapie was first described as a disease of sheep and goats over 250 years ago and demonstrated to be experimentally transmissible 50 years ago (Aguzzi & Polymenidou, 2004). There is no evidence that scrapie has ever transmitted to man. The only other known self-sustaining animal prion disease is chronic wasting disease in mule deer and elk in several states of the USA. Again there is no current evidence that this disease has transmitted to man.

BSE was first described in UK cattle in 1985 (Wells *et al.*, 1987) and is thought to have spread through oral consumption of ruminant-derived meat and bone meal (Wilesmith *et al.*, 1988; Brown, 1998). The disease spread widely, peaking in 1992 with over 180 000 clinical cases in the UK, although mathematical estimates suggest that 1–2 million cattle could have been infected but slaughtered and entered the human food chain before they were old enough to demonstrate evidence of clinical disease (Fig 1) (Anderson *et al.*, 1996). BSE has crossed into up to 20 other species, including domestic and exotic cats (Wyatt *et al.*, 1991; Kirkwood & Cunningham,

1994) and exotic ungulates in British zoos. In July 1988, the spread of BSE led the UK Government to restrict the use of ruminant-derived meat and bone meal as an animal feed and in November 1989 specified that bovine offals were banned for human consumption.

### Sporadic Creutzfeldt Jakob diseases

Sporadic CJD was the first described human prion disease, is of uncertain aetiology, has a worldwide distribution and an incidence of around one per million population per year (Will *et al.*, 1998). The median age at onset is around 68 years and the disease is characterised by a rapidly progressive dementia leading to death in around 4–6 months. The incidence of the disease varies with the codon 129 genotype of the *PRNP* gene, with 83% of patients homozygous for the expression of methionine at this locus (Deslys *et al.*, 1998). Molecular strain typing suggests that six forms of disease are dependent on codon 129 phenotype and strain of prion disease. One of the pathological hallmarks of sporadic CJD is the restriction of accumulation of plaques of prion protein to the central nervous system (CNS). However, with recently developed, more sensitive techniques, prion accumulation has also now been reported to be present in peripheral nerve (Favreux *et al.*, 2004) as well as in muscle, lymphoid tissue and olfactory epithelium (Glatzel *et al.*, 2003) at an advanced stage of clinical disease.

Although there are a small number of reports claiming transmission of sporadic CJD by inoculation of blood from patients with clinical disease into experimental rodents (Manuelidis *et al.*, 1985; Tateishi, 1985), these results have not been supported by further studies in primates (Brown *et al.*, 1994). Similarly, although there are a handful of reports of sporadic CJD arising after blood or plasma product transfusion (Klein & Dumble, 1993; Creange *et al.*, 1995, 1996; de Silva, 1996b; Patry *et al.*, 1998), in none of these has a causal link to a donor with CJD been established. Moreover a series of epidemiological case control (Kondo & Kuroiwa, 1982; Davanipour *et al.*, 1985; Harries-Jones *et al.*, 1988; Will, 1991; Wientjens *et al.*, 1996; Van Duijn *et al.*, 1998; Collins *et al.*, 1999), lookback (Esmonde *et al.*, 1993; Heye *et al.*, 1994; Operskalski & Morley, 1995) and surveillance (Evatt, 1998; Evatt *et al.*, 1998; Lee *et al.*, 1998) studies carried out over almost 25 years have failed to demonstrate evidence of transmission of sporadic CJD by blood components or plasma products. It seems likely therefore that the preclinical incubation period in sporadic CJD is sufficiently short, or peripheral blood infectivity is sufficiently low, as to make transmission of the disease by blood components and/or plasma products at worst a very rare event (de Silva & Mathews, 1993; Brown, 1995; Ricketts *et al.*, 1997; Will & Kimberlin, 1998).

Thus, although individuals suspected of having sporadic CJD are permanently deferred from blood donation, no other precautions, such as withdrawal of plasma products if the donor has contributed to the plasma pool, are undertaken.

This is because, although sporadic CJD is rare, a large number of donations would be expected to frequent withdrawal and plasma product donation.

### Familial human prion diseases

Familial human prion diseases are transmitted as autosomal dominant disorders, including Gerstmann-Sträussler-Eisenberg disease, familial insomnia and familial CJD, and are caused by mutations in the prion gene (*PRNP*). All forms of familial human prion disease transmit to other individuals with two or more blood transfusions, disease, or who have been advised that they are not to donate blood as a result of *PRNP* gene to pending, or past, blood donation as a precautionary measure.

### Acquired human prion diseases

The transmission of human prion disease was first reported in the Fore people of Papua New Guinea in the 1950s (Gajdusek & Zigas, 1960) and was shown to be transmitted during ritual cannibalism practices. The clinical features differ from those of sporadic CJD, with more prominent ataxia and longer incubation period. One time Fore ritual cannibalism was reported to have occurred in the 1960s and another in the 1970s. In the 1970s people and inter-tribe marriages were reported to have occurred around 1960, 1970 and 1980, and in the 1980s clinical disease was increasingly reported. The incubation period in prion disease is in the order of years. Inotropic transmission of CJD to experimental rodents (Manuelidis *et al.*, 1985; Tateishi, 1985) and direct inoculation of the CNS of experimental animals by surgical instruments (Bovine spongiform encephalitis material and converted into scrapie agent) have also occurred via experimental prion strains (Bovine spongiform encephalitis, agent transmitted by human to human (Bachmann *et al.*, 1991; Brown *et al.*, 1994). The presentation varies depending on the strain of prion, with a centrally transmitted case tending to have a longer incubation period of around 2 years, and developing progressive dementia remission of symptoms, and a peripherally transmitted case tend to have a shorter incubation period of around 17–47 years, and to develop ataxia and sensory disturbances (Tobler *et al.*, 1996).

### Variant CJD

Variant CJD was first described in Great Britain in 1996 as a result of systematic monitoring of the clinical phenotype of CJD in the UK by the National Surveillance Unit in Edinburgh, Scotland. It is unusual in that it presents with neurological features such as anxiety, depression and hallucinations, and does not develop progressive dementia, rapidity of progression, with an average clinical course of 14 months (Will *et al.*, 1998; Will & Mathews, 1993; Brown, 1995; Ricketts *et al.*, 1997; Will & Kimberlin, 1998).



Table 1. Iatrogenic transmission of Creutzfeldt-Jakob disease.

	Number	Incubation period (months)
Neurosurgical instruments	5	12–28
Intracerebral electrodes	2	16–20
Dura mater graft	120	18–216
Corneal graft	4	16–320
Human growth hormone	142	550–456
Human gonadotrophin	5	144–192

The incubation period for infections transmitted by peripheral inoculation is shorter than that when infection is directly in the brain (from Ironside and Head, 2003, with permission from Blackwell Publishing).

phalogram changes are observed, but magnetic resonance imaging (MRI) is more informative, with changes in the pulvinar ('pulvinar thalamic') in the majority of cases.

Neuropathologically, the disease is characterised by neural cell loss, a spongiosis and spongiform change with particularly florid amyloid plaques as a pathognomic feature (Fig 2) (Ironside & Head, 2003; Peden & Ironside, 2004). To date all

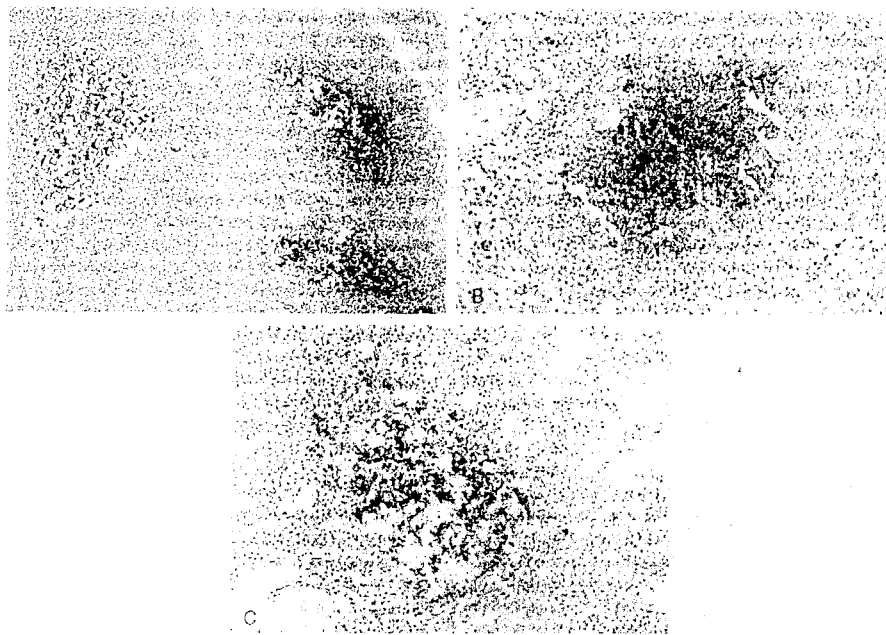


Fig 2. Immunocytochemistry for the prion protein (PrP) in lymphoid tissues in variant Creutzfeldt-Jakob disease shows staining of follicular dendritic cells and macrophages in (A) the tonsil, (B) spleen and (C) lymph node. Anti-PrP antibody (KG9) with haematoxylin counterstain [from Ironside and Head (2003) with permission from Blackwell Publishing].

clinical cases of variant CJD have occurred in methionine 129 homozygous individuals; it seems likely that valine homozygous and methionine/valine heterozygous individuals are more resistant to infection or, if infected, to the development of clinical variant CJD. In this context it may be relevant that methionine 129 human prion protein oligomerises more rapidly with beta-sheet formation whereas 129 valine tends to form alpha-helix rich monomers (Tahiri-Alaoui *et al.*, 2004). Furthermore it is of interest that following inoculation with prions, mice homozygous for human methionine developed 'typical' variant CJD, whilst those that were homozygous for valine appeared more resistant to infection and when this occurred, the clinical and pathological features were more similar to sporadic CJD (Wadsworth *et al.*, 2004). It is noteworthy, in this context that the second case of probable variant CJD prion transmission by blood transfusion was recorded in a methionine/valine heterozygous patient who did not develop clinical features of the disease despite surviving 5 years after transfusion (Peden *et al.*, 2004). This patient had been identified as part of the variant CJD lookback process and postmortem examination was requested following death from unrelated causes (*vide infra*).

Unlike sporadic and familial forms of CJD, patients with variant CJD show evidence of abnormal prion accumulation in follicular dendritic cells in peripheral lymphoid tissue including tonsils (Hill *et al.*, 1997b; Kawashima *et al.*, 1997), appendices, spleen (Hilton *et al.*, 1998) and lymph nodes (Hill *et al.*, 1999). In two patients, appendices removed 8 months and 2 years prior to the onset of clinical disease have also shown evidence of prion accumulation, although a sample removed 10 years prior to onset of clinical disease did not (Glatzel *et al.*, 2004).

The median age at death is 29 years (range 14–74 years) and has not altered over the first 10 years of the outbreak, suggesting an age-related susceptibility or exposure (Ghani *et al.*, 1998a; Boelle *et al.*, 2004). At the time of writing there have been 154 definite and probable cases of variant CJD in the UK, nine in France, two in Ireland and one in each of the USA, Canada, Italy, Saudi Arabia and Japan. In the UK, the incidence of clinical disease appears to have peaked around 2000 and has since fallen significantly (<http://www.cjd.ed.ac.uk>). However, although the outbreak thus far has been very much less than that which was initially feared (Cousens *et al.*, 1997; Ghani *et al.*, 1998b), with an upper boundary of around a further 70 new cases now predicted based on the pattern of clinical disease (Will, 2003; Smith *et al.*, 2004; Sneath, 2004), a recent retrospective study of tonsil and appendix samples demonstrated three of 12 500 samples positive for abnormal prion accumulation, suggesting that up to 3500 people could be infected with a prevalence of pre- or subclinical disease amongst the 10 to 30-year-old UK population of one of 10 000 (Hilton *et al.*, 2004). Ghani *et al.* (1998a) have suggested that up to 90% of individuals infected may have prolonged preclinical or true subclinical disease and that this could be related to codon 129 genotypes encoding valine homozygosity or methionine/valine heterozygosity. If transmissible prion infectivity is present in the peripheral blood of such asymptomatic individuals, the concern is that blood-derived products could provide a route to long-term persistence of variant CJD within the population.

#### Animal studies of peripheral blood infectivity and transmissibility

The route by which the prions disseminate and replicate following peripheral inoculation is of importance in understanding the likely distribution of infectivity and has been recently reviewed (Mabbott & Turner, 2005). Studies in knockout mice with deficiencies in PrP expression, or lacking various cellular compartments of their immune systems, have led to the conclusion that initial accumulation or replication in follicular dendritic cells is essential to peripheral transmission (McBride *et al.*, 1992; Bueler *et al.*, 1993; Fraser *et al.*, 1996; Brown *et al.*, 1997b; Klein *et al.*, 1997, 1998; Mabbott *et al.*, 1998). Indeed, infection and abnormal prion accumulation can be demonstrated in the lymphatic tissues of scrapie-infected rodents and sheep prior

to the stage of clinical disease in the central nervous system (Turner *et al.*, 1994; Mabbott *et al.*, 1998; Brown *et al.*, 1997b; Mabbott *et al.*, 1998). In sheep, the route of peripheral transmission is via the gut, with the prions entering the intestinal lumen and the infection spreading to the lymphatic system. The prions are thought to be taken up by follicular dendritic cells. A study of peripheral blood infectivity in sheep is reported by Ghani *et al.* (1998a) and the agent is transported by lymphocytes and macrophages and migratory follicular dendritic cells. Infection was first observed within 10 days after intramuscular inoculation of sympathetic and parasympathetic ganglia in the CNS. Agents in sympathetic ganglia were shown to interact with ganglionic acetylcholine receptors, a finding for this type of interaction has been considered a plausible route of peripheral transmission of prions from an infected sheep to the peripheral lymphatics.

As a potential route of peripheral transmission, it has been shown to be important in the transmission of scrapie from sheep to humans. The transmission of scrapie from sheep to humans is thought to have occurred in the 1970s and 1980s, with the first human case reported in 1976 (Cousens *et al.*, 1997; Ghani *et al.*, 1998b). It is estimated that these clinical cases were derived from the peripheral lymphatics of the prion carrier sheep.

In a recent retrospective study of tonsil and appendix samples infected with prions, the prions were found in peripheral lymphatic tissues, including the spleen, 10–15 times although the prevalence of prions in the peripheral lymphatics was low. The prevalence of prions in the peripheral lymphatics was low, suggesting that the general population of prion carriers is small (Brown *et al.*, 2004). The study also demonstrated that prions were found in the peripheral lymphatics of individuals who had been infected with prions approximately 10–15 years prior to the peripheral inoculation.

In sheep, naturally infected individuals are infected via oral and dermal routes, and the prions are transmitted to the peripheral lymphatics. The clinical incubation period for prions transmitted to sheep via the peripheral lymphatics is approximately 10–15 years, and the prions are found in the peripheral lymphatics of the prion carrier sheep.

#### Transmission of variant CJD by blood transfusion

The UK has a blood donor system which is based on the ability of the donor to donate blood. For example, individuals who are infected with variant CJD are not allowed to donate blood. However, the question arises as to whether individuals who are infected with variant CJD can donate blood. The UK has a blood donor system which is based on the ability of the donor to donate blood. For example, individuals who are infected with variant CJD are not allowed to donate blood. However, the question arises as to whether individuals who are infected with variant CJD can donate blood. The UK has a blood donor system which is based on the ability of the donor to donate blood. For example, individuals who are infected with variant CJD are not allowed to donate blood. However, the question arises as to whether individuals who are infected with variant CJD can donate blood.

The reverse arm of the surveillance scheme addresses the question as to whether any of the patients who have developed variant CJD could have become infected via a previous blood transfusion. The transfusion history of all patients developing variant CJD is assessed and the donors are traced and also flagged to the UK Office of National Statistics.

To date 17 variant CJD patients are known to have been blood donors (15 in the UK and two in France). Of the 50 recipients of blood components, 17 are still alive. Plasma from 23 donations was fractionated to produce albumin, immunoglobulin and clotting factor concentrates that were used in the UK, France, Belgium, Germany and Italy. In the UK it appears that the incidence of variant CJD peaked in about 2001 and is now declining (Fig 1).

To date there have been two cases of probable transmission of variant CJD prions via non-leucodepleted red cell concentrates. In the first episode, a 24-year-old individual gave a blood donation in 1996 (Llewelyn *et al.*, 2004). Three years later he developed variant CJD and died the subsequent year. The recipient of this donation in 1996 was aged 62 years and also received four other units of red cell concentrate to cover a surgical operation. In 2002 he became depressed and developed blurred vision, motor difficulties including a shuffling gait and cognitive impairment. An MRI of his brain was reported as normal. In 2003 he died of dementia. At autopsy, histology of his brain revealed characteristic features of variant CJD, and this was confirmed by proteinase-K resistance and typical features on Western blotting. Analysis of his *PRNP* gene revealed him to be homozygous for methionine at codon 129. A statistical assessment concluded that there was only a 1:15 000–1:30 000 chance of this occurring by coincidence.

A second individual was reported in 2004 as a result of the national surveillance of recipients of transfusions from donors who later developed variant CJD. This patient very likely became infected with variant CJD prions by a unit of red cell concentrate in 1999 from a donor who developed variant CJD 18 months later (Peden *et al.*, 2004). Although this patient died 5 years after the transfusion of unrelated causes with no clinical features of variant CJD, analysis of her lymphoid tissue at autopsy revealed that prion accumulation was present in the spleen and one cervical lymph node. There were no histological features or evidence of prion accumulation in her CNS. The other unusual feature as noted above, was that the *PRNP* gene was heterozygous at codon 129 for methionine/valine.

These two cases are therefore of great importance because they have demonstrated that variant CJD prions can be transmitted by blood transfusion from donors who are in a preclinical phase of disease at the time of donation and that methionine/valine heterozygous individuals can also be infected, although whether they are as susceptible to infection and/or the development of clinical disease as methionine homozygous individual remains uncertain (Aguzzi & Glatzel, 2004).

## Blood donor selection

Many countries have instituted policies of donor deferral for those who have spent time in the UK, France or more broadly Europe, based on the likely comparative level of risk with their indigenous population, the extent or pattern with which their population visit affected areas and the likely impact on their blood donor base.

In the UK, there are few epidemiological criteria that would allow identification of a 'high-risk' donor population. In response to the blood transfusion related transmissions of variant CJD, in 2004, a policy of deferral of donors who themselves have been recipients of blood components since 1980 was instituted to reduce the risk of tertiary or higher-order transmissions leading to a self-sustaining outbreak. This policy also has the advantage of reducing the risk of other blood borne infectious agents being recycled in the community by transfusion. There was concern that this would lead to a significant reduction in the donor base and that a sometimes precarious blood supply would be further compromised. Whilst about 5–10% donors have been lost from the UK blood donor panels, the impact has been mitigated by proactive recruitment campaigns to enlist more new donors.

## Importation of blood components

It is not likely to be feasible to import red cell or platelet concentrates due to the large volumes required, the short shelf life and lability of these components and concerns over the risk of other transmissible agents in some overseas donor populations. To reduce the risk of variant CJD transmission to children, in 2002 the decision was made to only use imported non-UK plasma to treat those born after 31 December 1995. This date was chosen because it was considered that BSE-infected foods had been largely eliminated from the diet by this date, and therefore, children born after this time were unlikely to be infected from food. In addition, with relatively small volumes of plasma, the product can be stored, transported frozen and be virus-inactivated.

## Donor screening

No immunological response to prion infection has yet been identified nor has DNA been found associated with disease transmission. Therefore, traditional serological and molecular biological approaches to donor screening are not currently feasible.

Several groups have looked at the possibility of using surrogate markers. The proteins 14-3-3 (Zerr *et al.*, 1998) and S100 (Otto *et al.*, 1998) are non-specific markers of CNS damage and are therefore likely to be elevated only in the clinical stages of disease. It has been shown that transcription of erythroid differentiation associated factor (EDAF) is depressed in the peripheral blood of animals suffering from prion disease (Miele *et al.*, 2001). The cause of this observation

is uncertain and it also currently remains unclear whether this could be translated into the setting of human clinical and preclinical disease and whether an appropriate differential exists between patients incubating variant CJD and normal individuals.

Infectivity has not thus far been detected in the peripheral blood of patients with clinical variant CJD by intracerebral inoculation into rodents despite the evidence of clinical transmission, reflecting the limitations of infectivity bioassays due to the species barrier and the small amounts of blood inoculated.

A central difficulty in the development of molecular assays is the differentiation of PrP<sup>Sc</sup> from PrP<sup>C</sup> (Minor, 2004). There are currently no monoclonal antibodies or other reagents of sufficient analytical specificity to differentiate between the normal and abnormal isoforms. Most assays therefore depend on differential physicochemical characteristics, such as resistance to proteinase-K digestion or display of additional or novel PrP epitopes following treatment with chaotropic agents, such as guanidine hydrochloride. The level of sensitivity required is challenging. Brown *et al.* (1999) has estimated that in the order 1 pg of PrP<sup>Sc</sup>/ml may be present in the peripheral blood of individuals in the pre- or subclinical phases of disease, in the context of around 100 ng/ml of PrP<sup>C</sup>, i.e. a ratio of 1 PrP<sup>Sc</sup> molecule:1 million PrP<sup>C</sup> molecules. There are also significant challenges in validating such assays. This would normally be undertaken using samples from individuals with the disease in question. However, there are very few patients alive at any one time with variant CJD and large amounts of blood cannot be drawn for ethical reasons. As it is not currently possible to determine who may, or may not, be incubating the disease, the assays will therefore need to be validated on brain homogenate-spiked human blood or animal endogenous infectivity samples posing questions around the extrapolation of the data to the human setting. Finally it should be borne in mind that it will not be possible to determine which of the donors with positive assays are actually incubating variant CJD and which of these are likely to go on to develop clinical disease. There is no treatment available at the present time to offer such individuals. There is concern, therefore, over the number of donors who may need to be deferred due to positive assay results and the potential impact of the introduction of such assays on the willingness of donors to donate (Blajchman *et al.*, 2004; McCullough *et al.*, 2004).

## Blood component processing

In October 1997, the UK Spongiform Encephalopathy Advisory Committee, advised that universal leucodepletion be considered. The UK Departments of Health commissioned an independent risk assessment by Det Norske Veritas Consulting (DNV) and asked the Blood Services to consider the feasibility (Comer & Spouge, 1999). Implementation was recommended in July 1998 and completed by the autumn of 1999 (Department of Health, 1998a,b). The measure was

predicated on studies suggesting that the infectivity of PrP<sup>Sc</sup> to be involved in a blood transfusion is contained in leucocytes were an important component of the infectivity in peripheral blood, supported by findings in experimental animal studies. Blood plasma was not considered to be infectious and is likely to reduce the potential infectivity by only about 4% (Frasse *et al.*, 2001; Brown *et al.*, 2001; St Romaine *et al.*, 2004). The goal was to reduce the risk of infection by only a small number of additional donors, considered to offer a number of additional units of blood reduction in transmission of all blood components (cytomegalovirus and human T-cell lymphotropic virus type 1) of alloimmunisation, immunomodulatory effects and transfusion-mediated graft versus host disease (Reddy *et al.*, 2004).

Other approaches under consideration to reduce the infection risk include the greater use of platelet concentrates from a single donor rather than a pool of many donors, extending the incubation period for platelet cryoprecipitate to all variants under the age of 10 years and further reduction in plasma volume in platelet concentrates.

Two components developing alternative methods to reduce the plasma contamination of blood components of magnitude, such as filtration, and would reduce the likelihood of transmission of variant CJD prions to the sub- or preclinical phase of disease, have been considered a significant challenge as studies undertaken naturally in other human populations have shown that therefore have been considered to be unlikely to be considered infected blood, thus raising the question of the validity of these models.

## Plasma product manufacture

In 1997, the Committee for Proprietary Medicines (Committee for Proprietary Medicines) and the licensing authority, recommended recall of factor VIII concentrate that had been manufactured from a plasma pool containing donations from two patients who subsequently developed variant CJD. The UK Department of Health Centre Doctors Organisation (DNV) advised that the recall was recommended that as variant CJD occurred in 1996 in the UK, it is likely that any risk of transmission would be reduced by using concentrates prepared from plasma donations collected in other countries, e.g. USA, where the incidence of variant CJD or PrP<sup>Sc</sup> (Llewelyn, 1997; Brown *et al.*, 2001) has been keen to try to evaluate the risk of blood transmission of variant CJD, so that the overall safety measure would be to recall in 2004, the original DNV risk assessment was updated to being further estimates of transmissibility. It has been suggested that animal studies using a prion bioassay to determine the infectivity of concentrates may be a more definitive method of determining the safety of the concentrate to be used. Some of the concerns that have been raised feature suggest that they will be predominantly of a theoretical